

Antimicrobial resistance of *Escherichia coli* isolated from Minas Frescal Cheese in the city of Rio de Janeiro -Phenotypic and genotypic profile

Resistência antimicrobiana em cepas de *Escherichia coli* isoladas de queijo Minas Frescal no município do Rio de Janeiro - Perfil fenotípico e genotípico

Bruna Amatto Duarte Pires^{I,*} D Juliana Wolff Salles de Oliveira^{II} Cristiane Rodrigues Silva^{II} Shirley de Mello Pereira Abrantes^I

Victor Augustus Marin[®] (D)

ABSTRACT

Introduction: Antimicrobial resistance is a public health problem. Foods may be carriers of antimicrobial resistant bacteria and resistance genes for humans. Objective: To isolate, identify, and evaluate the antimicrobial susceptibility and identification of resistance genes and integrons in Escherichia coli strains isolated from Minas Frescal cheese. Method: The presence of E. coli in thirty samples of Minas Frescal cheese was evaluated through the 3M™ Petrifilm plates. Thirty E. coli isolates were evaluated through the Disk Diffusion Susceptibility Test for 17 antimicrobial agents. Through PCR and Multiplex PCR the isolates were examined for the presence of 24 antimicrobial resistance genes. Genes and variable regions of class 1, 2 and 3 integrons were also evaluated. Results: Phenotypic resistance to one or more classes of antimicrobial agents were found in 50% of E. coli isolates. One hundred percent of the isolates showed PCR amplification for the *bla*_{TEM} gene and there was also amplification for *bla*_{SHV} and *tet*B genes. In addition, 46.6% of the phenotypically resistant isolates amplified to one or more classes of integrons. This is one of the first studies to identify these genes in Minas Frescal cheese in Brazil. Conclusions: Cheese can be a source of multiresistant bacteria and those can disseminate their resistance genes to other bacteria being in the food and human gastrointestinal tract, emphasizing the importance of Good Manufacturing Practices and greater supervision on products on sale.

KEYWORDS: Antimicrobial Resistance; E. coli; Integron; Minas Frescal Cheese

RESUMO

Introdução: A resistência antimicrobiana é um problema de saúde pública. Os alimentos podem ser veículos de bactérias resistentes a antimicrobianos e genes de resistência para os seres humanos. Objetivo: Isolamento, identificação, avaliação da susceptibilidade antimicrobiana e identificação de genes de resistência e integrons em Escherichia coli isoladas de queijo Minas Frescal. Método: Avaliação da presença de E. coli em trinta amostras de queijo Minas Frescal por meio de Placas Petrifilm 3M™. Trinta isolados de E. coli foram avaliados através do teste de sensibilidade por disco difusão para 17 agentes antimicrobianos. Isolados fenotipicamente resistentes foram examinados por meio de PCR e Multiplex PCR para 24 genes de resistência. Genes e regiões variáveis dos integrons de classe 1, 2 e 3 também foram avaliados. Resultados: Em 50% dos isolados verificou-se resistência fenotípica a uma ou mais classes de antimicrobianos. Cem por cento dos isolados apresentaram o gene bla_{TEM} e houve amplificação dos genes bla_{SHV} e tetB. Além disso, 46,6% dos isolados fenotipicamente resistentes amplificaram para uma ou mais classes de integrons. Este é um dos primeiros estudos a identificar esses genes em queijo Minas Frescal no Brasil. Conclusões: O queijo pode ser uma fonte de bactérias multirresistentes e estas podem disseminar seus genes de resistência a bactérias presentes no alimento e no trato gastrointestinal humano, demonstrando a importância das Boas Práticas de Fabricação e a necessidade de maior fiscalização dos produtos colocados à venda.

- Instituto Nacional de Controle de Qualidade em Saúde (INCQS), Fundação Oswaldo Cruz (Fiocruz), Rio de Janeiro, RJ, Brasil
- Universidade Federal do Estado do Rio de Janeiro (UNIRIO), Rio de Janeiro, RJ, Brasil
- * E-mail: amattobruna@gmail.com

Received: Apr 03, 2019 Approved: Jul 11, 2019

PALAVRAS-CHAVE: Resistencia Antimicrobiana; E. coli; Integron; Queijo Minas Frescal



INTRODUCTION

Food and its production may be vehicles for antimicrobial resistant bacteria and resistance genes toward humans and this fact is directly related to public health¹.

Antimicrobial resistant bacteria can spread antimicrobial resistance through horizontal gene transfer, a process that transfers resistance genes between bacteria². The acquisition and transfer of genes resistant to antimicrobial agents, associated with the selection caused by the intensive use of these substances in livestock, highlights the importance and need to evaluate the presence of antibiotic resistant bacteria in food³.

Interventions that limit the use of antibiotics in animal feed appear to be associated with a reduction in the development of antimicrobial resistance in humans and, therefore, further research is necessary to explore this complex association⁴.

Integrons play an important role in the acquisition and spread of antibiotic resistance genes⁵ and in multidrug resistance in Gram-negative bacteria because they capture and incorporate gene cassettes from the environment⁶. Thus, the detection and characterization of integrons containing antibiotic resistance genes are key steps to evaluate the potential of a given environment to represent a reservoir of antibiotic resistance⁷.

Antibiotic resistant bacteria and resistance genes can be transferred directly to humans via food chain⁸. Thus, it is important to know the phenotype and genes involved in antibiotic resistance of *Escherichia coli* strains that colonize food products, since these may act as reservoirs of antimicrobial resistance genes⁹.

It is also important to highlight that this is one of the first studies to evaluate the presence of these bacteria, their resistance genes and integrons in this food matrix, the Minas Frescal cheese.

METHODS

E. coli isolation

We evaluated 30 samples of Minas Frescal cheese from ten different brands, purchased in supermarkets in the city of Rio de Janeiro, Brazil.

For *E. coli* isolation, we used *E. coli* counting $3M^{\mathbb{M}}$ Petrifilm^{\mathbb{M}} plates and obtained several isolates. Then, we selected 30 *E. coli* isolates, each one from a different sample of Minas Frescal cheese. We checked and confirmed that all the isolates were strains of *E. coli* through polymerase chain reaction (PCR) according to Bej et al.¹⁰.

Kirby-Bauer disk diffusion sensitivity test

All 30 isolates of *E. coli* underwent the antimicrobial sensitivity test for 17 antimicrobial agents using the disk diffusion method in Mueller-Hinton agar, according to the criteria of the Clinical

and Laboratory Standards Institute (CLSI)¹¹. The antibiotics used in this test and their concentrations on the disks were: ampicillin (10 μ g), amoxicillin/clavulanic acid (20/10 μ g), ampicillin/sulbactam (10/10 μ g), tetracycline (30 μ g), cefoxitin (30 μ g), ceftriaxone (30 μ g), ceftazidime (30 μ g), cefepime (30 μ g), ciprofloxacin (5 μ g), nalidixic acid (30 μ g), imipenem (10 μ g), ertapenem (10 μ g), gentamicin (10 μ g), amikacin (30 μ g), streptomycin (10 μ g), sulfazotrim (30 μ g) and chloramphenicol (30 μ g).

Polymerase chain reaction

We tested all the isolates that presented phenotypic resistance to any of the 17 antimicrobial agents used in the sensitivity test for the presence of 24 antimicrobial resistance genes^{12,13,14,15,16,17,18,19} (Table 1).

Integron detection

We looked for the *intl* 1, *intl* 2, *intl* 3 genes (which encode for integrases classes 1, 2 and 3, respectively) and for the variable regions of the class 1 and 2 integrons by PCR (Table 2) in all the *E. coli* isolates according to Ryu et al.²⁰.

RESULTS

E. coli isolation in Minas Frescal Cheese

In a 1:10 dilution, most of the samples of Minas Frescal cheese (96.6%) presented a countless number of *E. coli* isolates. This fact demonstrates the level of contamination by this microorganism in this food. To obtain standard isolates of *E. coli*, that is, blue colonies with gas formation, we had to dilute the samples at 1:1,000 or 1:10,000.

Antimicrobial sensitivity test

Of the 30 isolates of *E. coli*, 15 (50%) were resistant to one or more antimicrobial classes (Table 3).

Resistance encoding genes

All 15 isolates of resistant *E. coli* tested positive for one or more resistance genes. All isolates (100%) amplified to the bla_{TEM} gene, and the isolate N also amplified to the bla_{SHV} gene and isolate C, for the *tet*B gene. This is one of the first studies to isolate these genes in Minas Frescal cheese.

Integrons

Of the phenotypically resistant isolates, 46.6% amplified to one or more classes of integrons. Isolate C amplified to class 2 and 3 integrons and also to the variable class 2 integron. Isolate H amplified to the class 3 integron and isolate M tested positive for class 1 and variable class 1 integron. Four other isolates (E, F, K, L) amplified only to the variable class 1 integron.



Table 1. Genes, sizes and references that were used for genetic identification of antimicrobial resistance.

Gene	Size (bp)	Reference			
СТХ	593	Al-Agamy et al., 2014 ¹²			
Multiplex: TEM/SHV/OXA	800/713/564	Dallene et al., 2010 ¹³			
Tet A/ Tet B/ Tet C/ Tet D/ Tet E	372/288/379/436/442	Srinivasan, 2008 ¹⁴			
Tet G	884	Ng, 2001 ¹⁵			
MOX	520				
СІТ	462				
DHA	405	C (1 1 · · · · · · · · · · · · · · · · ·			
EBC (MIR)	302	Székely et al., 2013 ¹⁶			
ACC	346				
FOX	190				
Multiplex: GES/ PER/ VEB	398/520/648	Al-Agamy et al., 2014 ¹²			
OXA48	438	Karuniawati et al., 2011 ¹⁷			
NDM	621	Karuniawati et al., 2011 ¹⁷			
КРС	635	Azimi et al., 2013 ¹⁸			
IMP	188 Zeighami et al., 2015				
VIM	390	Zeighami et al., 2015 ¹⁹			

bp: base pairs

 Table 2. Components used for integron identification

Gene	Primer	DNA Sequence (5'-3")	Size (bp)	Annealing Temperature (°C)=	
intl1	int1-L	CTGCGTTCGGTCAAGGTTCT	882	68	
	int1-F	GGAATGGCCGAGCAGATCCT	862		
intl2	int2.F	CACGGATATGCGACAAAAAGGT	746	59	
IIILIZ	int2.R	GTAGCAAACGAGTGACGAAATG	740	57	
intl3	int3.F	GCCTCCGGCAGCGACTTTCAG	939	62	
IIICIS	int3.R	ACGGATCTGCCAAACCTGACT	737	02	
Intl1 variable region	5′CS	GCCATCCAAGCAGCAAG	Variable	60	
Inter variable region	3′CS	AAGCAGACTTGACCTGA	variable	ΟU	
Intl2 variable region	hep74	CGGACGGCATGCACGATTTGTA	Variable	60	
	hep51	GATGCCATCGCAAGTACGAG	variable		

bp: base pairs

DISCUSSION

According to Amos et al.², class 1 integrons are vehicles for adaptive genes because of their ability to capture and integrate mobile cassettes into a variable region where they are expressed under a common promoter. These gene sequences can provide several phenotypes, including resistance to a wide range of antimicrobial classes.

Karuniawati, Saharman, Lestari¹⁷ described class 1 integron as the most frequent resistance gene carrier integron, and the production of the ß-lactamase enzyme is one of the most common resistance mechanisms. More recent studies on extended-spectrum ß-lactamase (ESBL) genes have shown that they are located in integrons. Therefore, ESBL genes can be located in integrons and easily transferred to various bacteria²¹.

Mohamed, Hassan, Ahmed²² evaluated class 1, 2, and 3 integrons and carbapenem resistance in Gram-negative bacteria and detected class 1 integron in most carbapenem resistant isolates. That is similar to what we found in this study, since of the three carbapenem (imipenem and ertapenem) resistant isolates (K, M, N), only one (N) did not amplify to that integron. The other two isolates (K and M) amplified to class 1 and/or variable class 1 integrons.

De Paula et al.²³ analyzed class 1, 2 and 3 integrons in Minas Frescal cheese samples and noticed that the presence of class 1



Table 3. Antimicrobial susceptibility test results according to the resistant E. coli isolates.

Isolate					Antibiotics	5				
А	Ampicillin									
В	Tetracylcine									
С	Ampicillin	Streptomycin	Amoxicillin/ clavulanic acid	Tetracylcine	Cefoxitin					
D	Ampicillin	Amoxicillin/ clavulanic acid	Cefoxitin							
E	Ampicillin	Streptomycin								
F	Ampicillin	Cefoxitin								
G	Ceftriaxone	Nalidixic acid	Cefepime	Ceftadizime						
Н	Ceftriaxone	Nalidixic acid								
I	Streptomycin	Nalidixic acid								
J	Nalidixic Acid									
К	Ampicillin	Imipenem								
L	Nalidixic Acid									
Μ	Streptomycin	Amoxicillin/ clavulanic acid	Ampicillin	Ampicillin / sulbactam	Cefoxitin	Ceftriaxone	Sulfazotrim	Nalidixic acid	Imipenem	Ertapenem
Ν	Amikacin	Streptomycin	Ampicillin	Tetracylcine	Cefoxitin	Ceftriaxone	Imipenem	Ertapenem	Cefepime	Ceftadizime
0	Ampicillin	Nalidixic acid								

and 2 integrons was 77% and 97% respectively. Class 3 integrons were not detected in any of the samples. These results are different from what we found in this study, in which we detected class 3 integron in 13.3% of the *E. coli* isolates from Minas Frescal cheese and class 1 and variable class 1 in 33.3% of the samples.

Regarding resistance to quinolone class, several mechanisms seem to contribute to this resistance. *Qnr* genes mediated by plasmids are a common mechanism of resistance to quinolone²⁴. A study suggests that the relation between the *qnr* gene and ESBL is associated with genes encoding $ESBL^{21}$. High levels of quinolone resistance in clinical strains of *E. coli* and other ESBL-producing bacteria were reported in several studies²⁵.

The studies described above corroborate our results, since the resistance to quinolone, represented by nalidyxic acid here, was always associated with the bla_{TEM} gene and, in two of the three isolates, we detected the integron and confirmed that the genes encoding for the ESBL may be located there.

The bla_{TEM} and bla_{SHV} genes are related to resistance to antimicrobial agents of the cephalosporin class. The SHV-type ESBL, together with the TEM-type, stood out during the 1980s as a cause of cephalosporin resistance among enterobacteriaceae²⁶, as occurs, for example, with isolate N, that contains the two genes (bla_{TEM} and bla_{SHV}) and is resistant to second, third and fourth generations cephalosporins (cefoxitin, ceftriaxone, ceftazidime, cefepime).

Traditionally, ESBL-producing isolates, mainly bla_{TEM} and bla_{SHV} producers, show co-resistance to aminoglycosides, tetracyclines and sulfonamides²⁷. This evidence corroborates the fact that

isolate N was also resistant to aminoglycosides (amikacin, streptomycin) and tetracycline.

The study of Juma et al.²⁸ evaluated the bla_{TEM} and bla_{SHV} genes in isolates of *E. coli* and found that bla_{TEM} was more common than bla_{SHV} . In this study, the presence of both genes (bla_{TEM} and bla_{SHV}) was confirmed in only one (6.6%) of the *E. coli* isolates (isolate M). The bla_{TEM} gene was found in all the other isolates (100%). A similar result was also found in the study of Freitas et al.²⁹.

Isolate C was resistant to ampicillin, streptomycin, tetracycline, cefoxitin and amoxicillin/clavulanic acid. Regarding the genotype, the following genes were found: bla_{TEM} , *tetB* and *IntI* 2, 3 and 2 variable. The resistance to tetracycline can, in this case, be confirmed by the existence of the *tetB* gene. Tetracyclines are the most widely used antimicrobials in animal therapy³⁰.

CONCLUSIONS

This study found *E. coli* in 100% of the commercial brand samples of Minas Frescal cheese. This fact is enough to show bacterial contamination from fecal origin in this type of food.

Half (50%) of the *E. coli* isolates were resistant to one or more antimicrobial classes and we were able to confirm by PCR the presence of the genes that justified the resistance phenotype presented by most of them. In addition, 46.6% of the phenotypically resistant isolates amplified to one or more integron classes, which corroborates the resistance phenotype presented by some isolates and demonstrates its importance in the spread of resistance genes among bacteria.



Since this is one of the first studies about Minas Frescal cheese in Brazil to detect the bla_{TEM} , bla_{SHV} tetB and intl 1, 2 and 3 genes, we expect it to contribute to the generation of data that improve the characterization of antibiotic resistant *E. coli* isolates and their spread through this type of food.

In the area of health surveillance, the studies about antimicrobials in food-producing livestock are very important, since

REFERENCES

- Caniça M, Manageiro V, Abriouel H, Moran-Gilad J, Franz CMAP. Antibiotic resistance in foodborne bacteria. Trends Food Sci Tech. 2019;84:41-4. https://doi.org/10.1016/j.tifs.2018.08.001
- Amos GCA, Ploumakis C, Zhang L, Hawkey PM, Gaze WH, Wellington EMH. The widespread dissemination of integrons throughout bacterial communities in a riverine system. ISME J. 2018;12(3):681-91. https://doi.org/10.1038/s41396-017-0030-8
- Cardoso P, Marin JM. Resistência antimicrobiana de isolados de Escherichia coli provenientes de queijo muçarela artesanal produzido no Brasil. Ars Vet. 2014;30(2):104-8. https://doi.org/10.15361/2175-0106.2014v30n2p104-108
- 4. Tang KL, Caffrey NP, Nóbrega DB, Cork SC, Ronksley PE, Barkema HW et al. Restricting the use of antibiotics in food-producing animals and its associations with antibiotic resistance in food-producing animals and human beings: a systematic review and meta-analysis. Lancet Planet Health. 2017;1(8):e316-27. https://doi.org/10.1016/S2542-5196(17)30141-9
- Guerin E, Cambray G, Sanchez-Alberola N, Campoy S, Erill I, Da Re S et al. The SOS response controls integron recombination. Science. 2009;324(5930):1034. https://doi.org/10.1126/science.1172914
- Ding J, Zhuochang Chen Z, Li Y, Zhang Q, Li X. Detection of integrons in *Escherichia coli* producing plasmid-mediated AmpC B-lactamases. Int J Clin Exp Med. 2019;12(2):1690-6.
- Yaqoob M, Wang LP, Fang T, Cheng-Ping LU. Occurrence and transmission of class 1 and 2 integrons among phenotypic highly ampicillinresistant avian *Escherichia coli* isolates from Pakistan. World J Microbiol Biotech. 2011;27(9):2041-50. https://doi.org/10.1007/s11274-011-0666-x
- Jiang X, Shi L. Distribution of tetracycline and trimethoprim sulfamethoxazole resistance genes in aerobic bacteria isolated from cooked meat products in Guangzhou, China. Food Control. 2013;30(1):30-4. https://doi.org/10.1016/j.foodcont.2012.06.042
- Jouini A, Ben Slama K, Sáenz Y, Klibi N, Costa D, Vinué L et al. Detection of multiple-antimicrobial resistance and characterization of the implicated genes in *Escherichia coli* isolates from foods of animal origin in Tunis. J Food Prot. 2009;72(5):1082-8.
- 10. Bej AK, Dicesare JL, Haff L, Atlas RM. Detection of *Escherichia coli* and *Shigella* spp. in water by using the

this is the main way through which bacteria from animal GI tract acquire resistance. Furthermore, studies about Good Manufacturing Practices in the production of this type of food are also important.

We also expect that this study can work as a foundation for further studies and public health policies so that food will not become a reservoir or transmitter of multiresistant bacteria to people.

polymerase chain reaction and gene probes for uid. Appl Environ Microbiol. 1991;57(4):1013-7.

- Clinical and Laboratory Standards Institute CLSI. Performance standards for antimicrobial susceptibility testing. Wayne: Clinical and Laboratory Standards Institute; 2015.
- Al-Agamy MH, Shibl AM, Ali MS, Khubnani H, Radwan HH, Livermore DM. Distribution of B-lactamases in carbapenem-non-susceptible *Acinetobacter baumannii* in Riyadh, Saudi Arabia. J Glob Antimicrob Resist. 2014;2(1):17-21. https://doi.org/10.1016/j.jgar.2013.08.004
- Dallenne C, Costa A, Decre D, Favier C, Arlet G. Development of a set of multiplex PCR assays for the detection of genes encoding important B-lactamases in Enterobacteriaceae. J Antimicrob Chemother. 2010;65(3):490-5. https://doi.org/10.1093/jac/dkp498
- 14. Srinivasan V, Nam H, Sawant AA, Headrick SI, Nguyen LT, Oliver SP. Distribution of tetracycline and streptomycin resistance genes and class 1 integrons in Enterobacteriaceae isolated from dairy and nondairy farm 293 soils. Microb Ecol. 2008;55(2):184-93. https://doi.org/10.1007/s00248-007-9266-6
- Ng LK, Martin I, Alfa M, Mulvey M. Multiplex PCR for the detection of tetracycline resistant genes. Mol Cell Probes. 2001;15(4):209-15. https://doi.org/10.1006/mcpr.2001.0363
- 16. Székelya E, Damjanovac I, Jánváric LE, Vas K, Molnár S, Bilca DV et al. First description of blaNDM-1, blaOXA-48, blaOXA-181 producing Enterobacteriaceae strains in Romania. Int J Med Micro. 2013;303(8):697-700. https://doi.org/10.1016/j.ijmm.2013.10.001
- 17. Karuniawati A, Saharman YR, Lestari DC. Detection of carbapenemase encoding genes in enterobacteriace, *Pseudomonas aeruginosa*, and *Acinetobacter baumanii* isolated from patients at intensive Care Unit Cipto Mangunkusumo Hospital in 2011. Acta Med Indones. 2013;45(2):101-6.
- Azimi L, Lari AR, Talebi M, Namvar AE. Soleymanzadeh-Moghadam S. Evaluation of phenotypic methods for detection of *Klebsiella Pneumoniae* carbapenemase-producing *K. Pneumoniae* in Tehran. J Med Bacteriol. 2013;2(3-4):26-31.
- Zeighami H, Haghi F, Hajiahmadi F. Molecular characterization of integrons in clinical isolates of betalactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* in Iran. J Chemother. 2015;27(3):145-51. https://doi.org/10.1179/1973947814Y.0000000180



- 20. Ryu SH, Park SG, Choi SM, Hwang YO, Ham HJ, Kim SU et al. Antimicrobial resistance and resistance genes in *Escherichia coli* strains isolated from commercial fish and seafood. Int J Food Microbiol. 2012;152(1-2):14-8. http://doi.org/10.1016/j.ijfoodmicro.2011.10.003
- 21. Hadizadeh M, Norouzi A, Taghadosi R, Mohebi S, Mohammadi M, Hasanzadeh A et al. Prevalence of qnr, intl, and intll genes in extendedspectrum beta-lactamase (ESBL)-producing *Escherichia coli* isolated from clinical samples in Iran. Trop J Pharm Res. 2017;16(1):141-7. https://doi.org/10.4314/tjpr.v16i1.18
- Mohamed MSE, Hassan AT, Ahmed SMK. Prevalence of class 1, 2, and 3 integrons and carbapenem resistance in Gram-negative bacteria. Egyptian J Med Micro. 2016;25(3):67-73. https://doi.org/10.12816/0036812
- Paula ACL, Medeiros JD, Azevedo AC, Chagas JMA, Silva VL, Diniz CG. Antibiotic resistance genetic markers and integrons in white soft cheese: aspects of clinical resistome and potentiality of horizontal gene transfer. Genes. 2018;9(2). http://doi.org/10.3390/genes9020106
- 24. Nordmann P, Poirel L. Emergence of plasmid-mediated resistance to quinolones in Enterobacteriaceae. J Antimicrob Chemother. 2005;56(3):463-9. http://doi.org/10.1093/jac/dki245
- 25. Jiang Y, Zhou Z, Qian Y, Wei Z, Yu Y, Hu S et al. Plasmid-mediated quinolone resistance determinants qnr and aac (6')-Ib-cr in extended-spectrum

B-lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* in China. J Antimicrob Chemother. 2008;61(5):1003-6. http://doi.org/10.1093/jac/dkn063

- Pitout JD. Extraintestinal pathogenic Escherichia coli: an update on antimicrobial resistance, laboratory diagnosis and treatment. Expert Rev Anti Infect Ther. 2012;10(10):1165-76. http://doi.org/10.1586/eri.12.110
- Morosini MI, Castillo MG, Coque TM, Valverde A, Novais A, Loza E et al. Antibiotic coresistance in extended-spectrum-lactamase-producing Enterobacteriaceae and in vitro activity of tigecycline. Antimicrob Agents Chemother. 2006;50(8):2695-9. https://doi.org/10.1128/AAC.00155-06
- Juma BW, KAriuki S, Waiyaki PG, Mutug MM, Bulimo WD. The prevalence of TEM and SHV genes among extended-spectrum beta-lactamase-producing *Klebsiella pneumoniae* and *Escherichia coli*. Afr J Pharmacol Ther. 2016;5(1):1-7.
- Freitas ALPD, Machado DP, Soares FDSC, Barth AL. Beta-lactamases de espectro ampliado em *Klebsiella* spp e em *Escherichia coli* obtidas em um hospital escola brasileiro: detecção, prevalência e tipagem molecular. Brazil J Microbiol. 2003;34:344-8.
- Schwarz S, Chaslus-Dancla E. Use of antimicrobials in veterinary medicine and mechanisms of resistance. Vet Res. 2001;32(3-4):201-25. https://doi.org/10.1051/vetres:2001120

Acknowledgements

We thank the Brazilian Institute for Health Quality Control (INCQS/FIOCRUZ) and the Federal University of the State of Rio de Janeiro (UNIRIO) for the use of their research laboratories. This study was done with the support of the Brazilian Coordination for the Improvement of Higher Education Personnel (CAPES) - Financing Code 001 and the Carlos Chagas Filho Research Foundation of the State of Rio de Janeiro (FAPERJ).

Conflict of Interest

Authors have no potential conflict of interest to declare, related to this study's political or financial peers and institutions.



This publication is licensed under the Creative Commons Attribution 3.0 Unported license. To view a copy of this license, visit http://creativecommons.org/licenses/by/3.0/deed.pt.