

Norovirus in foods

Norovírus em alimentos

Isabelle da Silva Luz*

Marize Pereira Miagostovich

ABSTRACT

Introduction: Noroviruses (NoV) are important causative agents of foodborne gastroenteritis outbreaks associated with the consumption of fruits, leafy vegetables, bivalve molluscs and delicatessen foods. The establishment of laboratory surveillance networks in different continents has demonstrated increased epidemiological importance of those viruses. In Brazil, the NoV infection is considered an important public health issue with socioeconomic burden, but the investigation of these viruses in foodborne outbreaks is still restricted to research laboratories. NoV infections have become more known especially with the consolidation of the cruise ship market in the country since 2004. **Objective:** This study aims to present advances related to NoV research in foods, highlighting features of this pathogen and strategies for its detection in these matrices. **Method:** An integrative review, collecting scientific articles with the objective of dealing with the main aspects of NoV, was carried out. **Results:** A broad literature review was performed, describing the main results in the literature and discussing aspects such as foodborne diseases, viruses as food contaminants, stability and disinfection, foodborne outbreaks associated with NoV, food associated with NoV contamination, NoV concentration and detection methods in food, risk assessment studies and prevention and control. **Conclusions:** records of foodborne outbreaks associated with NoV and the increasing genetic diversity of these viruses reinforce the need for laboratory and epidemiological surveillance, especially in developing countries, such as Brazil.

KEYWORDS: Norovirus; Foods; Disease Outbreaks; Gastroenteritis; Methods; Sanitary Surveillance

RESUMO

Introdução: Os norovírus (NoV) são importantes agentes causadores de gastroenterite de origem alimentar, com surtos associados ao consumo de frutas, vegetais folhosos, moluscos bivalves e alimentos de *delicatessen*. O aumento da importância epidemiológica destes vírus tem sido demonstrado pelo estabelecimento de redes laboratoriais de vigilância em diversos continentes. As infecções por NoV se tornaram mais conhecidas especialmente com a consolidação do mercado de navios de cruzeiros no país a partir de 2004. **Objetivo:** Este estudo tem como objetivo apresentar avanços relacionados à pesquisa de NoV em alimentos, destacando características deste patógeno e estratégias para sua detecção nestas matrizes. **Método:** Foi realizada uma revisão integrativa, pelo levantamento de artigos científicos com o objetivo de tratar dos principais aspectos de NoV. **Resultados:** Foi realizada uma ampla revisão da literatura, com a descrição dos principais resultados presentes na literatura consultada e a discussão de aspectos como doenças transmitidas por alimentos (DTA), vírus como contaminantes de alimentos, estabilidade e desinfecção, surtos de origem alimentar associados aos NoV, alimentos associados à contaminação por NoV, métodos de concentração e detecção de NoV em alimentos, estudos de avaliação de risco e prevenção e controle. **Conclusões:** Os registros de envolvimento de NoV em surtos de origem alimentar e a crescente diversidade genética destes vírus reforçam a necessidade de vigilância laboratorial e epidemiológica sobretudo nos países em desenvolvimento, como o Brasil.

¹ Fundação Oswaldo Cruz, Instituto Oswaldo Cruz, Laboratório de Virologia Comparada e Ambiental, Rio de Janeiro, RJ, Brasil

* E-mail: isabelle.luz@ioc.fiocruz.br
belleluz@gmail.com



INTRODUCTION

First described in the 1970s with the use of electronic immunomicroscopy¹, Norwalk viruses, as noroviruses (NoV) were then known, had their epidemiological importance recognized only from the 1990s onward with the emergence of molecular techniques of cloning and nucleotide sequencing, which allowed the production of diagnostic supplies^{2,3,4,5,6}. Today, NoV are recognized as the main agents that cause outbreaks and sporadic cases of acute non-bacterial gastroenteritis in humans^{7,8,9,10}.

The impact of NoV infections in industrialized countries is evident in the number of epidemiological surveillance networks established on different continents. Electronic platforms such as Noronet (Netherlands), Foodborne Viruses in Europe (European Union), Norovirus Outbreak Reporting Tool (England), Episurv (New Zealand), OzFoodNet (Australia), Calicinet and National Outbreak Reporting System (United States) have been promoting inter-laboratory integration by sharing epidemiological and molecular data on outbreaks, providing information on genotype circulation and the emergence of new variants^{11,12,13,14}. In these countries, where the diagnosis is well established, NoV are responsible for more than 200,000 deaths/year, mainly of children under 5 years of age⁹.

In Brazil, people became more aware of NoV infections after the consolidation of the cruise ship market in the country in the 2004/2005 season (<http://www.abremar.com.br/download/fgv2015.pdf>), once gastroenteritis outbreaks are common in these settings^{10,15,16,17,18}. Furthermore, several studies conducted in the country have demonstrated the impact of NoV infections in different populations, including outbreaks, sporadic cases, hospitalized patients, as well as the occurrence of asymptomatic infections and cases associated with persistent diarrhea^{19,20,21,22,23,24,25,26,27,28,29,30,31,32}. The environmental dissemination of NoV in different water matrices in the country has also been demonstrated with concentrations reaching $1.5E + 04 - 0.3E + 05$ (GC)/L in samples of crude sewage^{33,34,35,36,37,38,39,40}.

Concerning foodborne gastroenteritis, which belong to the group of foodborne diseases (FBD), NoV was associated with 38 outbreaks (0.9%) out of a total of 9,719 cases reported by the Ministry of Health (MS) in relation to foodborne gastroenteritis, in the period of 2000-2014⁴¹. In over 10,000 outbreaks of gastroenteritis associated with food contamination reported in recent years, more than half did not have a defined etiologic agent. Therefore, as in most countries, the determination of foodborne outbreaks associated with NoV is based on epidemiological investigations and laboratory tests performed on clinical specimens of individuals involved in these outbreaks⁴².

The detection of human NoV in food is hampered by the complexity of the food matrix and by the presence of low levels of virus particles, which results in outbreak underreporting^{43,44}. This review aims to present NoV as the main viral agents associated with outbreaks of foodborne gastroenteritis, describing their general characteristics and the progress related to the research of these viruses in food matrices.

METHOD

This study, prepared as an integrative review, was conducted according to the methodology described by Sobral and Campos⁴⁵ for the survey of scientific articles to address the main aspects of NoV caused by the consumption of contaminated food, as well as related infections. The research of scientific literature was carried out on the PubMed database (using keywords like: norovirus on foods, methodologies for norovirus on foods, norovirus review) and the database of the Brazilian Ministry of Health.

RESULTS AND DISCUSSION

Foodborne Diseases (FBD)

FBD is a generic term applied to a syndrome usually consisting of anorexia, nausea, vomiting and/or diarrhea, with or without fever, attributed to the ingestion of contaminated food or water. However, digestive symptoms are not the only manifestations of these diseases, since extraintestinal infections in different organs and systems may also occur, according to the agent involved. In addition to bacteria and toxins, FBD can also be caused by toxic substances, parasites, and viruses⁴⁶.

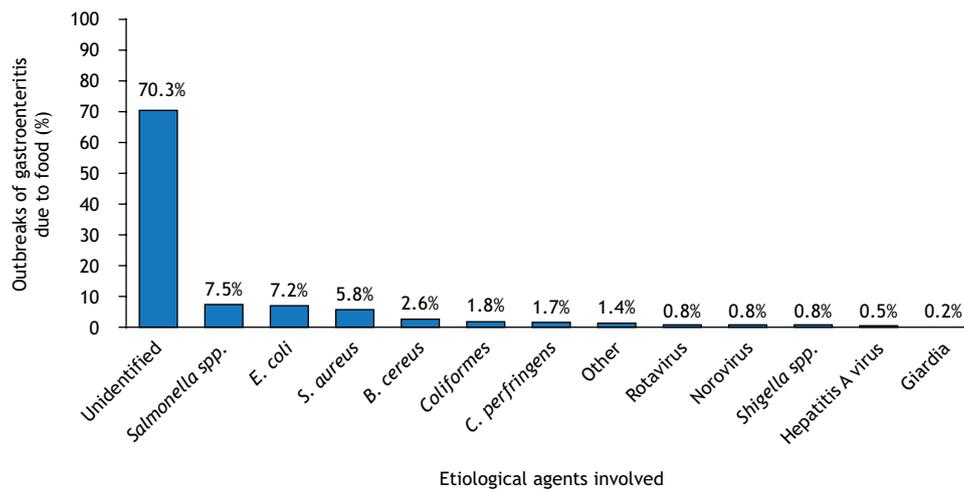
The epidemiological profile of FBD in Brazil is still poorly understood. Only a few states and/or municipalities have statistical information and data on the most common etiologic agents, food items frequently implicated, the highest risk population and contributing factors^{46,47}. There are also cases of FBD that are not notified to health authorities, since many foodborne pathogens cause mild symptoms and patients do not always seek medical help⁴⁸.

In many countries, including Brazil, the description of outbreaks (in which two or more people have a similar disease after ingesting food and/or water from the same source) is restricted to those involving a larger number of people or when the duration of the symptoms is longer⁴⁹.

The Figure shows the main etiologic agents identified in the outbreaks of FBD that occurred in Brazil between 2007 and 2016, highlighting the number of unidentified agents and the small number of cases associated with NoV, as well as other viral agents (rotavirus and hepatitis A virus).

Viruses as food contaminants

As fecal-oral transmission viruses, human enteric viruses are important contaminants of water and food, mainly because they are non-enveloped viruses that are resistant to adverse conditions, both in the human body (stomach acidity) and in the environment⁵⁰. Like NoV, hepatitis A (HAV) and E (HEV) viruses, enterovirus, astrovirus, parvovirus, rotavirus, adenovirus (AdV) 40 and 41, and, more rarely, coronaviruses are also associated with foodborne infections^{51,52}.



Source: Adapted from Ministry of Health (2016)⁴⁷; *data subject to updates.

Figure. Outbreaks of foodborne gastroenteritis identified in Brazil in the period 2007-2016* according to the etiological agent involved.

Although the stability of these viruses in different matrices depends on several environmental factors, such as pH, heat and resistance to cleaning agents, the low NoV infectious dose (18 virus particles can cause disease) represents a relevant factor in the transmissibility of these viruses^{16,53,54,55,56,57}. Ingestion of contaminated water or food is the main route of infection in these cases, however, the associated disease may occur indirectly from contact with contaminated fomites⁵⁸.

An important factor to be considered in the transmission of NoV is the large number of asymptomatic infections^{59,60,61}. NoV outbreaks often involve food preparation by a handler in the food service environment, where direct hand or gloved hand contact and inadequate cleaning are identified as common contributing factors⁶². Virus dispersal by food handlers has the potential for contamination in these environments, where large amounts of food are prepared in relatively small areas, involving the interaction of several employees⁶³.

Norovirus

Belonging to the *Norovirus* genus, *Caliciviridae* family, NoV are a group of non-enveloped, icosahedral viruses, of approximately 27 to 38 nm in diameter, named after the Greek word *calyx* (chalice), referring to the depressions of this format on the surface of the virus^{64,65}. These viruses were previously referred to by other names, such as small round-structured viruses and Norwalk-like viruses⁶⁶.

The genome consists of a single-stranded positively polarized RNA ranging from 7.3 to 7.5 kb, arranged in three open reading frames (ORF) and with a poly (A) tail at the end 3'^{2,3,67}. ORF1 encodes a polyprotein, which is cleaved in at least six non-structural proteins, including RNA-dependent RNA polymerase (RdRp); ORF2 and ORF3 respectively encode the VP1 and VP2 proteins of the viral capsid⁶⁸.

Because of the genetic diversity of the genus, NoV are classified into genogroups (G) and genotypes (GG) by nucleotide sequencing of the complete genomic region coding for the VP1 capsid

protein⁶⁹. Today, NoV have been classified into seven genogroups (GI-GVII)⁷⁰ of which three (GI, II and IV) infect humans⁷¹.

NoV GII.4 has been associated with most outbreaks and sporadic cases worldwide, mainly due to the emergence of new variants that become dominant at intervals of 2 to 3 years⁷². However, in 2013, GII.P17 appeared as a new genotype with evolution potential similar to that of GII.4, changing the epidemiology of NoV in the world⁷³. The antigenic drift and recombination of hotspot, mainly from the ORF1/ORF2 junction region, have been reported as an important mechanism for the evolution of NoVs, leading to the emergence of new viruses^{74,75,76,77,78,79,80}. A number of recombinant NoV strains have already been described, so that analysis of more than one region of the genome may be important for the detection of single or recombinant strains⁸¹.

In Brazil, the genetic diversity of NoV was demonstrated by the detection of different genotypes of human genogroups GI (GI.1-4, GI.7-8), GII (GII.1-9, GII.12-17, GII. 20-22, GII.b, GII.g, GII.e) and IV (GIV.1), as well as GII.4 and recombinant variants (US95_96, Kairo_2003, Asia_2003, Hunter_2004, Yerseke_2006a, Den Haag_2006b, New Orleans_2009 and Sydney_2012)^{77,82,83,84,85,86}.

NoV infection in humans is characterized as a self-limited gastrointestinal infection with symptoms including nausea, vomiting, diarrhea, malaise, abdominal pain, muscle aches, anorexia, headache and low fever. Symptoms usually begin 1 to 2 days after consumption of contaminated food or water and persist for 1 to 8 days⁶⁴.

Outbreak investigations have implicated vomiting as a route of transmission through inhaled aerosols or the direct contamination of surfaces^{87,88,89}. The infection affects all age groups, occurring mainly in domestic and institutional environments, such as hospitals, schools, restaurants, nursing homes and sea cruises^{60,65,90,91}. The epidemiology of NoV is complex and influenced by many factors, including population immunity, virus evolution, seasonality, virus stability in the environment and the frequent occurrence of asymptomatic infections^{59,60, 61,92,93,94,95}.



Stability and disinfection

NoV remain infectious after treatment with commonly used disinfectants, such as alcohols and quaternary ammonium compounds, as well as after heating at a temperature of 60° C for 30 minutes, 20% ether for 18 hours at 4° C and when exposed to pH 2.7 for three hours at room temperature⁹⁶. They may also be stable to inactivation after treatment with 3.75 to 6.25 mg/L chlorine (free residual chlorine of 0.5 to 1.0 mg/L), the concentration that is found in water supply systems. However, NoV particles are inactivated after treatment with 10 mg/L chlorine. Studies have shown that NoV are more resistant to chlorine inactivation than poliovirus type 1, human rotavirus (Wa), rotavirus simion (SA11) and bacteriophage F2⁹⁷.

According to Mormann et al.⁹⁸, measures used by the food processing industry for preservation purposes and processes used by consumers for preparation and storage would be sufficient to inactivate NoV in contaminated food. Therefore, the validation of thermal inactivation conditions in specific foods is necessary⁹⁹.

Considering the stability of NoV in the environment, Baert et al.¹⁰⁰ have developed a review on the efficacy of preservation methods used for virus inactivation in food. The authors suggested that food preservation methods such as heating, hydrostatic high pressure processing and irradiation are more effective in inactivating pathogens than freezing, refrigeration, reduced water activity, acidification or modified atmosphere packaging. They also highlighted the time-temperature combination and the variable efficacy of sanitizers on the food matrix in relation to viral strains.

The unavailability of cell strains for replication of human NoV in the laboratory resulted in the use of viruses belonging to the same genus as substitutes for predicting the behavior of NoV in food stability studies. Because they share similar structural and genetic characteristics and propagate in cell culture, murine norovirus-1 (MNV-1) (genogroup V) has been used in these studies^{101,102}. Also included are canine calicivirus (CaCV) used by Rutjes et al.^{101,102} on lettuce and cream samples, and Tulane virus (TV), a calicivirus belonging to the *Recovirus*^{93,104} genus. A study by Wang et al.⁹⁴ demonstrated that MNV-1, TV and HAV may be resistant on the surface of alfalfa seeds for an extended period (22° C for up to 50 days). These viruses could contaminate shoots after germination and be carried to irrigation water.

FBD outbreaks associated with NoV

According to a survey of the literature on global epidemiological trends from outbreaks from 1983 to 2011, Matthews et al.¹⁰⁵ observed that the majority of NoV infections were transmitted by food source routes (54%), with person-to-person transmission coming next (26%). However, this was a meta-analysis of published outbreaks and not necessarily based on population-based surveillance data. Furthermore, attack rate (defined as number of cases per exposed person) and distribution of genotypes are relevant factors for the investigation of outbreaks¹⁰⁶.

To estimate the proportion of foodborne infections caused by NoV on a global scale, Verhoef et al.¹⁴ used multiple international outbreak surveillance systems (NoroNet, Calicinet, Episurv) and systematic review of the literature. They demonstrated that

although the proportion of outbreaks caused by NoV GII.4 was smaller than that associated with other genotypes, the absolute contribution of foodborne outbreaks by NoV GII.4 to the social and economic costs caused by this virus is considerable.

Food associated with contamination by NoV

Fresh food subject to environmental contamination and handling¹⁰⁷, like fruit, leafy vegetables¹⁰⁸ and bivalve molluscs¹⁰⁹ is most at risk of NoV contamination. These types of food, in addition to being eaten raw, are subject to considerable human handling and undergo industrial sanitary treatments that do not guarantee the total elimination of pathogens¹¹⁰. Deli and ready-to-eat items that do not undergo further processing, like cold sandwiches^{111,112}, vegetable salads¹¹³ and confectionery products¹¹⁴, are also commonly associated with outbreaks.

Fruit and leafy vegetables

Outbreaks related to various types of products, including fresh cut fruits, lettuce, tomatoes, melons, salads, chives, strawberries, raspberries and parsley were associated with human NoV^{115,116,117}. Several outbreaks involved in the consumption of fresh produce were known or suspected of contamination in the field, suggesting that irrigation water can be a route of contamination^{93,118}.

Previous research with hydroponic lettuce cultivation has shown that viruses can be internalized through the root and disseminated to the aerial parts of the plant^{93,119}. Plant growth medium has been shown to play a significant role in the internalization of the pathogen by uptake by the root system¹²⁰.

In the United States, human NoV accounts for more than 40% of diseases related to fresh produce each year^{121,122} and, according to the European Food Safety Authority (EFSA) and the European Center for Prevention and Control Disease Control (ECDC), 11.6% of cases of viral infections were caused by consumption of vegetables, fruit, berries, juices and mixed foods in 32 European countries in the year 2013^{123,124}.

Bivalve molluscs

Bivalve molluscs are classically known for their high risk of microbiological contamination, since they are natural accumulators of particles dispersed in water. Bacteriological parameters have been used as a food safety regulatory criterion to evaluate the contamination of these food items, as well as of their culture water, especially after events of potential fecal contamination¹²⁵.

However, concentrations of *Escherichia coli* and coliforms in oysters and culture water may be reduced within a few days due to inactivation and elimination under environmental and tidal influences. However, this does not occur with viruses¹²⁶. A characteristic of outbreaks related to this source is its frequent association with multiple strains of virus observed in both infected patients and in the food involved¹²⁷.

Most of NoV outbreaks associated with bivalve molluscs are linked to the consumption of oysters because they are commonly



eaten raw, although some outbreaks have been linked to cooked oysters¹²⁸. Previous studies have also reported that imported frozen oysters were associated with outbreaks of gastroenteritis by NoV in Australia and in the United States^{129,130}.

Oyster cleansing tanks have been used to reduce bacterial contamination, however, standard cleansing procedures are ineffective for viral contaminants, as demonstrated by the high NoV levels detected in commercially distributed oysters in Italy and in the United States^{131,132}. In artificially contaminated and cleansed oysters, human AdV were detected up to 168 h and MNV-1 up to 96 h of cleansing, with viral quantification ranging from $3.2E + 05$ CG/g to $4.4E + 07$ CG/g for AdV and $3.5E + 04$ CG/g to $2.9E + 06$ CG/g for MNV-1 after 14 days of analysis¹³³.

A study carried out in the United Kingdom in 2011 showed that 76.2% ($n = 844$) of samples collected in the oyster production areas showed positive results for NoV GI and/or GII¹³⁴. In an outbreak of gastroenteritis associated with oyster consumption in a restaurant, also in the UK, NoV GI and GII were detected at concentrations < 100 copies/g (the theoretical limit of detection of the assay is 13 copies/g of the sample's digestive gland) and 1,736 copies/g, respectively¹³⁵.

In Brazil, GI NoV were detected in *Crassostera gigas* cultivated in marine farms for 14 days, with concentrations of $1.2 E + 06$ CG/g, and GII NoV in sea water, with concentrations of $7.5 E + 13$ CG/g¹³⁶. In subsequent investigations, NoV were not found by Souza et al.¹³³ in naturally contaminated oysters.

Deli items and ready-to-eat food

A method for detection of NoV was evaluated by Stals et al.¹³⁷ in ready-to-eat food items like penne salad, soups, sandwiches and compound meals, finding that the recovery of GI and GII NoV was influenced by the level of viral inoculum and the type of food. Furthermore, MNV-1 was successfully evaluated as process control by the same detection methodology.

In a gastroenteritis outbreak caused by NoV, Malek et al.¹³⁸ found that consumption of meat from a deli resulted in 137 sick persons on 13 independent rafting trips for a period of one month. The same virus sequence was found in fecal samples obtained from persons who participated of five different trips.

In Brazil, NoV GI.1 was identified in a sample of butter with herbs and NoV GII.4 in naturally contaminated cheese and white sauce samples, related to an outbreak of acute gastroenteritis on a cruise ship¹⁵. Also in this study, partial sequencing of the RNA polymerase gene showed the presence of GII.4 strains, confirming previous studies describing the incidence and distribution of this genotype in the world⁷⁴, including Brazil^{21,139}.

Methods of concentration and detection of NoV in foods

According to Baert et al.¹¹⁴, three food categories are considered when choosing concentration and virus detection methodologies: food rich in water and carbohydrates (fruits and vegetables); rich in protein and fat (ready-to-eat) and bivalve molluscs, due

to the accumulation and concentration of viral particles and other pathogens in the digestive system⁶⁷.

The steps required for the detection of viruses in these matrices include 1) virus concentration and purification, 2) nucleic acid extraction, 3) detection, and 4) confirmation¹⁴⁰. Concentration of viral particles to a smaller sample volume is the most critical step of the process and is particularly necessary because of the low levels of virus that may occur in the matrices^{137,141,142}. During concentration of the virus, molecules such as polysaccharides, proteins and fatty acids are removed to prevent inhibition of subsequent RNA extraction and molecular detection^{143,144}.

Elution-concentration protocols, based on the recovery of viral particles from the food surface using an appropriate buffer followed by concentration of the eluted viruses, include polyethylene glycol (PEG) precipitation, ultracentrifugation, ultrafiltration, immunoconcentration and cation separation. Different methodologies have viral recovery rates influenced by the concentration of inoculum and the type of food analyzed¹³⁷.

The efficiency of these methods has been evaluated in several studies with the aim of providing information on viral recovery. In a study by Summa et al.¹⁴⁵, lettuce, ham and raspberry samples were artificially contaminated with GII NoV, for comparison of four viral recovery methods based on ultrafiltration techniques, immunomagnetic separation, ultracentrifugation and PEG precipitation. Ultracentrifugation produced higher recovery efficiencies in lettuce and ham, while PEG precipitation generated higher NoV recovery yields in raspberries.

Other methods, initially described for NoV concentration from different water matrices, have been adapted for recovery of these viruses in food matrices. The use of common methods for different matrices may be useful in the investigation of outbreaks, in which samples of various origins are available. The negatively charged membrane filtration concentration method described for recovery of NoV from sea water¹⁴⁶ was adapted for samples of fresh lettuce and minas cheese by the direct elution of these food items^{147,148,149}.

The organic flocculation method using skim milk¹⁵⁰ was also successfully adapted for virus recovery from strawberries¹⁵¹. When compared to PEG precipitation methods and filtration with negatively charged membranes, it showed recovery percentage of 2.5 and 32 times higher than the other methodologies, respectively. Organic flocculation is a low cost method, since it uses only one step in the concentration of the samples, saving time and reagents. The Table summarizes the viral recovery rates obtained with these methodologies in studies conducted in Brazil.

RNA extraction is the second step in the NoV detection strategy. Extraction protocols involve (1) lysis of the viral capsid and (2) isolation of RNA⁶⁷. However, direct viral RNA extraction techniques involve treatment of the food product by viral elution with a reagent based on guanidine/phenol isothiocyanate, followed by purification of the extracted RNA. Direct RNA extraction was applied to food composed of protein and/or fat, with 1 to 10^2 units of NoV



Table. NoV recovery efficiencies in food.

Viral concentration methods	Food samples	Average NoV recovery efficiency (%)	References
Filtration with negatively charged membranes	Cheese	6.0-56.3	147
	Lettuce	5.2-72.3	
Filtration with negatively charged membranes	Lettuce	3.5-32.0	149
Filtration with negatively charged membranes	Lettuce	0.06-0.67	148
Organic flocculation	Strawberry	1.29-41.37	151

detected, which were recovered in 10 g to 30 g of hamburger, turkey, roasted beef, penne, tagliatelle and deli ham^{114,152,153}.

The first detailed description of the use of molecular methodologies for understanding foodborne outbreaks was described in the United States after the detection of NoV in contaminated ham¹¹¹. Molecular methods of reverse transcription followed by polymerase chain reaction (RT-PCR) are used for the detection and quantification of NoV. The RT-qPCR quantitative method, which incorporates a fluorescently labeled probe or fluorescently colored dye specifically interleaved into the reaction mixture, has been most recommended because of its sensitivity, specificity and speed¹⁵⁴.

However, this methodology based on a standard curve requires careful calibration and offers relative quantification with inter-laboratory variations¹⁵⁵. As the detection of small viral concentrations is the rule for food matrices, the interpretation of results should follow well-established criteria¹⁴⁰.

Despite sensitivity, the molecular assay has limitations because it does not provide infectivity data; and detected RNA may come from an integral viral particle or be a residual molecule¹⁵⁶. Recently, procedures for the pretreatment and/or use of dyes that interleave in RNA and DNA, such as propionate monoazide (PMA) in molecular methodologies, have been used for the detection and determination of infectivity of human NoV^{157,158}, with amplification occurring only in viral genomes of whole particles, i.e. infectious particles^{159,160}.

Another important issue in detecting NoV from food matrices is the use of viruses as internal process control. MNV-1, Mengov (strain MC₀), feline calicivirus (FCV), and bacteriophages such as MS2 and PP7^{148,161} are examples of viruses that have been successfully used^{135,162,163,164,165,166}, with bacteriophages being more readily available for laboratory production of food microbiology¹⁶⁷.

After viral detection, another fundamental step is the molecular characterization of NoV by genome nucleotide sequencing. Complete sequencing of ORF2 that encodes the viral capsid VP1 protein (1,600 base pairs) is the standard for the molecular characterization of genotypes and phylogenetic studies⁷¹. However, partial sequencing of this region of the genome has been used for rapid characterization of genotypes by the use of primers targeting smaller regions of ORF2, designated C (5' end of ORF2) and D (3' end of ORF2)^{168,169,170}.

For the molecular characterization of variants of GII.4, Vega et al.¹⁷¹ have developed an amplification protocol that uses

primers that target the coding region of the P2 subdomain of the VP1 protein of the viral capsid, since most of the mutations that differentiate genotypes and variants occur in that region. Today, the molecular characterization of NoV is enabled by the National Institute for Public Health and the Environment (RVMI), which provides the automatic genotyping tool by the insertion of nucleotide sequences of the genome in this platform¹⁷².

In 2013, Technical Specifications (TS) developed by the European Committee for Standardization [(ECS)/TC 275/WG 6] and approved by the International Organization for Standardization (ISO) established standardized methodologies for detection of NoV and HAV (ISO/TS 15216 -1, 2013, and ISO/TS 15216-2, 2013) into high-risk food categories. It was a significant advance in food virology studies⁵¹.

Risk assessment studies

Quantitative microbial risk assessment (QMRA) has become a valuable tool for characterizing risks of foodborne disease associated with pathogens. Nevertheless, a substantial share of the studies are related to bacterial agents^{173,174,175}. Regarding NoV, QMRA models were developed to evaluate the NoV risk in drinking water¹⁷⁶ and recreation water^{177,178}. In food, QMRA studies for NoV are limited and concentrated on the initial contamination of fresh produce^{179,180,181}.

A review of microbiological risk assessment studies on water and safety of fresh products revealed that viruses had higher risk estimates compared to bacterial agents. Leafy vegetables were identified as the products of greatest concern when compared to other foodstuffs¹⁸².

However, a study by Stals et al.¹¹² presented a quantitative model of exposure to NoV focusing on the potential transmission during the preparation of sandwiches. They found that a single dispersion of NoV per food handler could cause mean levels of 43±18, 81±37 and 18±7 NoV particles in the sandwiches, hands and work surfaces, respectively.

Prevention and control

Rapid laboratory diagnosis is an important tool for targeting NoV outbreak control through the choice of appropriate intervention and control practices, such as cleaning and disinfection protocols, isolation, patient grouping based on symptoms, exclusion of symptomatic employees or food handlers or, ultimately, establishment closure¹⁸³.



Contamination control of food, water, surfaces and fomites, as well as the proper hygiene of food handlers, is essential to reduce transmission rates⁶⁵. In the case of infected food handlers, absence is recommended for at least 3 days after the end of the symptoms. Infected adults and children should be kept out of school and work for the same period of time. In case of outbreaks, the operations of cruise ships, resorts, campgrounds and restaurants should be discontinued in order to avoid exposure of a new susceptible population¹⁸⁴. Contaminated surfaces after episodes of vomiting or diarrhea should be disinfected with 5% -25% or 1,000 to 5,000 ppm hypochlorite solution¹⁸⁵.

The increasing clinical significance of human NoV infections suggests the need for an effective vaccine that would promote blockade of transmission pathways particularly for high-risk populations such as food handlers, military personnel, the elderly, children and immunodeficient individuals, thereby improving food safety, public health and biodefense⁴².

The development of vaccines for NoV has been directed to the expression of viral capsid proteins like virus-like particles (VLPs) in different vectors^{97,186,187}. A broad-coverage bivalent vaccine that uses VLPs from a consensus of three NoVGII.4 variants in combination with NoVGI.1 is in the final stages of testing by the Takeda Vaccines group^{188,189,190}. Despite the advances already achieved, one of the major challenges in vaccine creation is the great genetic variability of these viruses and the replacement of pandemic strains in short time intervals, as observed for influenza A virus¹⁹¹.

REFERENCES

1. Kapikian AZ, Wyatt RG, Dolin R, Thornhill TS, Kalica AR, Chanock RM. Visualization by immune electron microscopy of a 27-nm particle associated with acute infectious nonbacterial gastroenteritis. *J Virol.* 1972;10(5):1075-81.
2. Jiang X, Graham DY, Wang K, Estes MK. Norwalk virus genome cloning and characterization. *Science.* 1990;250(4987):1580-3.
3. Jiang X, Wang M, Wang K, Estes MK. Sequence and genomic organization of Norwalk virus. *Virology.* 1993;195(1):51-61. <https://doi.org/10.1006/viro.1993.1345>
4. Ando T, Mulders MN, Lewis DC, Estes MK, Monroe SS, Glass RI. Comparison of the polymerase region of small round structured virus strains previously classified in three antigenic types by solid-phase immune electron microscopy. *Arch Virol.* 1994;135(1-2):217-26. <https://doi.org/10.1007/BF01309781>
5. Hardy ME, Estes MK. Completion of the Norwalk virus genome sequence. *Virus Genes.* 1996;12(3):287-90. <https://doi.org/10.1007/BF00284649>
6. Atmar RL, Estes MK. Diagnosis of noncultivable gastroenteritis viruses, the human caliciviruses. *Clin Microbiol Rev.* 2001;14(1):15-37. <https://doi.org/10.1128/CMR.14.1.15-37.2001>
7. Scallan E, Hoekstra RM, Angulo FJ, Tauxe RV, Widdowson MA, Roy SL, et al. Foodborne illness acquired in the United States - major pathogens. *Emerg Infect Dis.* 2011;17(1):7-15. <https://doi.org/10.3201/eid1701.P11101>
8. Rodríguez-Lázaro D, Cook N, Ruggeri FM, Sellwood J, Nasser A, Nascimento MS, et al. Virus hazards from food, water and other contaminated environments. *FEMS Microbiol Rev.* 2012;36(4):786-814. <https://doi.org/10.1111/j.1574-6976.2011.00306.x>
9. Ahmed SM, Hall AJ, Robinson AE, Verhoef L, Premkumar P, Parashar UD, et al. Global prevalence of norovirus in cases of gastroenteritis: a systematic review and meta-analysis. *Lancet Infect Dis.* 2014;14(8):725-30. [https://doi.org/10.1016/S1473-3099\(14\)70767-4](https://doi.org/10.1016/S1473-3099(14)70767-4)
10. Robilotti E, Deresinski S, Pinsky BA. Norovirus. *Clin Microbiol Rev.* 2015;28(1):134-64. <https://doi.org/10.1128/CMR.00075-14>
11. Duizer E, Kroneman A, Siebenga J, Verhoef L, Vennema H, Koopmans M et al. Typing database for noroviruses. *Eurosurveill.* 2008;13(19):1-2.
12. Yu Y, Cai H, Hu L, Lei R, Pan Y, Yan S, et al. Molecular epidemiology of oyster-related human noroviruses and their global genetic diversity and temporal-geographical distribution from 1983 to 2014. *Appl Environ Microbiol.* 2015;81(21):7615-24. <https://doi.org/10.1128/AEM.01729-15>
13. Tavoschi L, Severi E, Niskanen T, Boelaert F, Rizzi V, Liebana E et al. Food-borne diseases associated with frozen berries consumption: a historical perspective, European Union, 1983 to 2013. *Eurosurveill.* 2015;20(29):21193. <https://doi.org/10.2807/1560-7917.ES2015.20.29.21193>



14. Verhoef L, Hewitt J, Barclay L, Ahmed SM, Lake R, Hall AJ, et al. Norovirus genotype profiles associated with foodborne transmission, 1999-2012. *Emerg Infect Dis*. 2015;21(4):592-9. <https://doi.org/10.3201/eid2104.141073>
15. Gabbay YB, Siqueira, JAM, Lima ICG, Teixeira DM, Aragão GC, Barbagelata LS, et al. Norovirus outbreak in a cruise ship along the Brazilian coast, March 2011. *Rev Pan-Amaz Saúde*. 2011;5(1):43-51. <https://doi.org/10.5123/S2176-62232014000100005>
16. Morillo SG, Luchs A, Cilli A, do Carmo Sampaio Tavares Timenetsky M. Rapid detection of norovirus in naturally contaminated food: foodborne gastroenteritis outbreak on a cruise ship in Brazil, 2010. *Food Environ Virol*. 2012;4(3):124-9. <https://doi.org/10.1007/s12560-012-9085-x>
17. Bert F, Scaioli G, Gualano MR, Passi S, Specchia ML, Cadeddu C, et al. Norovirus outbreaks on commercial cruise ships: a systematic review and new targets for the public health agenda. *Food Environ Virol*. 2014;6(2):67-74. <https://doi.org/10.1007/s12560-014-9145-5>
18. Morillo SG, Luchs A, Cilli A, Ribeiro CD, Carmona RCC, Timenetsky MCST. Norovirus GII. Pe genotype: tracking a foodborne outbreak on a cruise ship through molecular epidemiology, Brazil, 2014. *Food Environ Virol*. 2017;9(2):142-8. <https://doi.org/10.1007/s12560-016-9272-2>
19. Victoria M, Carvalho-Costa FA, Heinemann MB, Leite JP, Miagostovich M. Prevalence and molecular epidemiology of noroviruses in hospitalized children with acute gastroenteritis in Rio de Janeiro, Brazil, 2004. *Pediatr Infect Dis J*. 2007;26(7):602-6. <https://doi.org/10.1097/INF.0b013e3180618bea>
20. Soares CC, Santos N, Beard RS, Albuquerque MC, Maranhão AG, Rocha LN et al. Norovirus detection and genotyping for children with gastroenteritis, Brazil. *Emerg Infect Dis*. 2007;13(8):1244-6. <https://doi.org/10.3201/eid1308.070300>
21. Nakagomi T, Correia JB, Nakagomi O, Montenegro FM, Cuevas LE, Cunliffe NA et al. Norovirus infection among children with acute gastroenteritis in Recife, Brazil: disease severity is comparable to rotavirus gastroenteritis. *Arch Virol*. 2008;153(5):957-60. <https://doi.org/10.1007/s00705-008-0060-7>
22. Campos GS, Moreau VH, Bandeira A, Barberino G, Almeida PF, Amador DM et al. Molecular detection and genetic diversity of norovirus in hospitalized young adults with acute gastroenteritis in Bahia, Brazil. *Arch Virol*. 2008;153(6):1125-9. <https://doi.org/10.1007/s00705-008-0078-x>
23. Ribeiro LR, Giuberti RS, Barreira DM, Saick KW, Leite JP, Miagostovich MP et al. Hospitalization due to norovirus and genotypes of rotavirus in pediatric patients, state of Espírito Santo. *Mem Inst Oswaldo Cruz*. 2008;103(2):201-6. <https://doi.org/10.1590/S0074-02762008000200013>
24. Ferreira MS, Xavier MP, Fumian TM, Victoria M, Oliveira SA, Pena LH et al. Acute gastroenteritis cases associated with noroviruses infection in the state of Rio de Janeiro. *J Med Virol*. 2008;80(2):338-44. <https://doi.org/10.1002/jmv.21059>
25. Andreasi MS, Cardoso D, Fernandes SM, Tozetti IA, Borges AM, Fiaccadori FS et al. Adenovirus, calicivirus and astrovirus detection in fecal samples of hospitalized children with acute gastroenteritis from Campo Grande, MS, Brazil. *Mem Inst Oswaldo Cruz*. 2008;103(7):741-4. <https://doi.org/10.1590/S0074-02762008000700020>
26. Ferreira MS, Victoria M, Carvalho-Costa FA, Vieira CB, Xavier MP, Fioretti JM et al. Surveillance of norovirus infections in the state of Rio de Janeiro, Brazil 2005-2008. *J Med Virol*. 2010;82(8):1442-8. <https://doi.org/10.1002/jmv.21831>
27. Barreira DM, Ferreira MS, Fumian TM, Checon R, de Sadovsky AD, Leite JP et al. Viral load and genotypes of noroviruses in symptomatic and asymptomatic children in Southeastern Brazil. *J Clin Virol*. 2010;47(1):60-4. <https://doi.org/10.1016/j.jcv.2009.11.012>
28. Ferreira MS, Cubel Garcia RC, Xavier MP, Ribeiro RL, Assis RM, Mota MC et al. Genotyping of gastroenteric viruses in hospitalized children: first report of norovirus GII.21 in Brazil. *Mem Inst Oswaldo Cruz*. 2012;107(8):1064-7. <https://doi.org/10.1590/S0074-02762012000800017>
29. Ferreira MS, Xavier MP, Tinga AC, Rose TL, Fumian TM, Fialho AM et al. Assessment of gastroenteric viruses frequency in a children's day care center in Rio de Janeiro, Brazil: a fifteen year study (1994-2008). *PLoS One*. 2012;7(3):e33754. <https://doi.org/10.1371/journal.pone.0033754>
30. Raboni SM, Damasio GA, Ferreira CE, Pereira LA, Nogueira MB, Vidal LR et al. Acute gastroenteritis and enteric viruses in hospitalised children in southern Brazil: aetiology, seasonality and clinical outcomes. *Mem Inst Oswaldo Cruz*. 2014;109(4):428-35. <https://doi.org/10.1590/0074-0276140066>
31. Oliveira DMM, Souza M, Fiaccadori FS, Santos HCP, Cardoso DDP. Monitoring of Calicivirus among day-care children: evidence of asymptomatic viral excretion and first report of GI.7 Norovirus and GI.3 Sapovirus in Brazil. *J Med Virol*. 2014;86(9):1569-75. <https://doi.org/10.1002/jmv.23791>
32. Amaral MS, Estevam GK, Penatti M, Lafontaine R, Lima IC, Spada PK et al. The prevalence of norovirus, astrovirus and adenovirus infections among hospitalized children with acute gastroenteritis in Porto Velho, state of Rondônia, western Brazilian Amazon. *Mem Inst Oswaldo Cruz*. 2015;110(2):215-21. <https://doi.org/10.1590/0074-027610140381>
33. Miagostovich MP, Ferreira FF, Guimarães FR, Fumian TM, Diniz-Mendes L, Luz SL et al. Molecular detection and characterization of gastroenteritis viruses occurring naturally in the stream waters of Manaus, central Amazonia, Brazil. *Appl Environ Microbiol*. 2008;74(2):375-82. <https://doi.org/10.1128/AEM.00944-07>
34. Victoria M, Guimarães FR, Fumian TM, Ferreira FF, Vieira CB, Shubo T et al. One year monitoring of norovirus in a sewage treatment plant in Rio de Janeiro, Brazil. *J Water Health*. 2010;8(1):158-65. <https://doi.org/10.2166/wh.2009.012>
35. Prado T, Silva DM, Guilayn WC, Rose TL, Gaspar AM, Miagostovich MP. Quantification and molecular characterization of enteric viruses detected in effluents from two hospital wastewater treatment plants. *Water Res*. 2011;45(3):1287-97. <https://doi.org/10.1016/j.watres.2010.10.012>



36. Vieira CB, Mendes AC, Guimarães FR, Fumian TM, Leite JP, Gaspar AM et al. Detection of enteric viruses in recreational waters of an urban lagoon in the city of Rio de Janeiro, Brazil. *Mem Inst Oswaldo Cruz.* 2012;107(6):778-84. <https://doi.org/10.1590/S0074-02762012000600012>
37. Moresco V, Viancelli A, Nascimento MA, Souza DS, Ramos AP, Garcia LA, et al. Microbiological and physicochemical analysis of the coastal waters of southern Brazil. *Mar Pollut Bull* 2012;64(1):40-8. <https://doi.org/10.1016/j.marpolbul.2011.10.026>
38. Prado T, Guilayn WC, Gaspar AM, Miagostovich MP. The efficiency of concentration methods used to detect enteric viruses in anaerobically digested sludge. *Mem Inst Oswaldo Cruz.* 2013;108(1):77-83. <https://doi.org/10.1590/S0074-02762013000100013>
39. Victoria M, Fumian TM, Rocha MS, Dalmao F, Leite JP, Girones R et al. Gastroenteric virus dissemination and influence of rainfall events in urban beaches in Brazil. *J Appl Microbiol.* 2014;117(4):1210-18. <https://doi.org/10.1111/jam.12592>
40. Prado T, Gaspar AM, Miagostovich MP. Detection of enteric viruses in activated sludge by feasible concentration methods. *Braz J Microbiol.* 2014;45(1):343-9. <https://doi.org/10.1590/S1517-83822014000100049>
41. Ministério da Saúde (BR), Secretaria da Vigilância em Saúde, Coordenação Geral de Doenças Transmissíveis. *Vigilância epidemiológica das doenças transmitidas por alimentos - VE-DTA.* São Paulo: Ministério da Saúde; 2014.
42. Dicaprio E, Ma Y, Hughes J, Li J. Epidemiology, prevention, and control of the number one foodborne illness: human norovirus. *Infect Dis Clin North Am.* 2013;(27):651-74. <https://doi.org/10.1016/j.idc.2013.05.009>
43. Le Guyader FS, Neill FH, Dubois E, Bon F, Loisy F, Kohli E et al. A semiquantitative approach to estimate Norwalk-like virus contamination of oysters implicated in an outbreak. *Int J Food Microbiol.* 2003;87(1-2):107-12. [https://doi.org/10.1016/S0168-1605\(03\)00058-8](https://doi.org/10.1016/S0168-1605(03)00058-8)
44. Li D, Baert L, Xia M, Zhong W, Jiang X, Uyttendaele M. Effects of a variety of food extracts and juices on the specific binding ability of norovirus GII.4 P particles. *J Food Prot.* 2012;75(7):1350-4. <https://doi.org/10.4315/0362-028X.JFP-12-002>
45. Sobral FR, Campos CJG. Utilização de metodologia ativa no ensino e assistência de enfermagem na produção nacional: revisão integrativa. *Rev Esc Enferm USP.* 2012;46(1):208-18. <https://doi.org/10.1590/S0080-62342012000100028>
46. Ministério da Saúde (BR), Secretaria de Vigilância em Saúde. *Manual integrado de prevenção e controle de doenças transmitidas por alimentos.* Brasília, DF: Secretaria de Vigilância em Saúde; 2010.
47. Ministério da Saúde (BR), Secretaria de Vigilância em Saúde, Unidade de Vigilância das Doenças de Transmissão Hídrica e Alimentar. *Surtos de doenças transmitidas por alimentos no Brasil.* Brasília, DF: Secretaria de Vigilância em Saúde; 2016.
48. Forsythe SJ. *Microbiology of safe food.* 2a ed. Oxford: Willey-Blackwell; 2010.
49. Carmo GMI, Oliveira AA, Dimech CP, Santos DA, Almeida MG, Berto LH et al. *Vigilância epidemiológica das doenças transmitidas por alimentos no Brasil, 1999 -2004.* Bol Eletr Epidemiol. 2005;5(6):1-7.
50. Cuellar JL, Meinhoefel F, Hoehne M, Donath E. Size and mechanical stability of norovirus capsids depend on pH: a nanoindentation study. *J Gen Virol.* 2010;91:2449-56. <https://doi.org/10.1099/vir.0.021212-0>
51. Hennechart-Collette C, Martin-Latil S, Guillier L, Perelle S. Determination of which virus to use as a process control when testing for the presence of hepatitis A virus and norovirus in food and water. *Int J Food Microbiol.* 2015;2(202):57-65. <https://doi.org/10.1016/j.ijfoodmicro.2015.02.029>
52. Richards GP, Cliver DO, Greening GE. Foodborne viruses. In: Salfinger Y, Tortoello ML. *Compendium of methods for the microbiological examination of foods.* 2nd ed. Washington, DC: American Public Health Association; 2015. Chapter 44.
53. Appleton H. Control of food-borne viruses. *Br Med Bull.* 2000;56(1):172-83. <https://doi.org/10.1258/0007142001902879>
54. Koopmans M, Bonsdorff CH, Vinjé J, Medici D, Monroe S. Foodborne viruses. *FEMS Microbiol Rev.* 2002;26(2):187-205. <https://doi.org/10.1111/j.1574-6976.2002.tb00610.x>
55. Koopmans M, Duizer E. Foodborne viruses: an emerging problem. *Int J Food Microbiol.* 2004;90(1):23-41. [https://doi.org/10.1016/S0168-1605\(03\)00169-7](https://doi.org/10.1016/S0168-1605(03)00169-7)
56. Bozkurt H, D'Souza DH, Davidson PM. Thermal inactivation kinetics of human norovirus surrogates and hepatitis A virus in turkey deli meat. *Appl Environ Microbiol.* 2015;14:4850-9. <https://doi.org/10.1128/AEM.00874-15>
57. Lee M, Seo DJ, Seo J, Oh H, Jeon SB, Ha SD et al. Detection of viable murine norovirus using the plaque assay and propidium-monoazide-combined real-time reverse transcription-polymerase chain reaction. *J Virol Methods.* 2015;221:57-61. <https://doi.org/10.1016/j.jviromet.2015.04.018>
58. Knight A, Haines J, Stals A, Li D, Uyttendaele M, Knight A et al. A systematic review of human norovirus survival reveals a greater persistence of human norovirus RT-qPCR signals compared to those of cultivable surrogate viruses. *Int J Food Microbiol.* 2016;216:40-9. <https://doi.org/10.1016/j.ijfoodmicro.2015.08.015>
59. Phillips G, Tam CC, Rodrigues LC, Lopman B. Prevalence and characteristics of asymptomatic norovirus infection in the community in England. *Epidemiol Infect.* 2010;138(10):1454-8. <https://doi.org/10.1017/S0950268810000439>
60. Nicolay N, McDermott R, Kelly M, Gorby M, Prendergast T, Tuite G et al. Potential role of asymptomatic kitchen food handlers during a food-borne outbreak of norovirus infection, Dublin, Ireland, March 2009. *Eurosurveill.* 2011;16(30):pii: 19931.
61. Jeong AY, Jeong HS, Lee JS, Park YC, Lee SH, Hwang IG et al. Occurrence of norovirus infections in asymptomatic food handlers in South Korea. *J Clin Microbiol.* 2013;51(2):598-600. <https://doi.org/10.1128/JCM.01856-12>



62. Centers for Disease Control and Prevention (CDC). Multisite outbreak of norovirus associated with a franchise restaurant -Kent County, Michigan, May 2005. *MMWR*. 2006;55(14):395-7.
63. World Health Organization, Food and Agriculture Organization of the United Nations, Codex Alimentarius. Guidelines on the application of general principles of food hygiene to the control of viruses on food. Rome: FAO; 2012[acesso 31 jan 2016]. (CAC/GL 79-2012). Disponível em: http://www.codexalimentarius.org/download/standards/13215/CXG_079e.pdf
64. Grove SF, Lee A, Lewis T, Stewart CM, Chen H, Hoover DG. Inactivation of foodborne viruses of significance by high pressure and other processes. *J Food Prot*. 2006;69(4):957-68. <https://doi.org/10.4315/0362-028X-69.4.957>
65. Glass RI, Parashar UD, Estes MK. Norovirus gastroenteritis. *N Engl J Med*. 2009;361(18):1776-85. <https://doi.org/10.1056/NEJMra0804575>
66. Lambden PR, Caul EO, Ashley CR, Clarke IN. Sequence and genome organization of a human small round-structured (Norwalk-like) virus. *Science*. 1993;259(5094):516-9. <https://doi.org/10.1126/science.8380940>
67. Stals A, Mathijis E, Baert L, Botteldoorn N, Denayer S, Mauroy A et al. Molecular detection and genotyping of noroviruses. *Food Environ Virol*. 2012;4(4):153-67. <https://doi.org/10.1007/s12560-012-9092-y>
68. Gao Z, Liu B, Huo D, Yan H, Jia L, Du Y et al. Increased norovirus activity was associated with a novel norovirus GII.17 variant in Beijing, China during winter 2014-2015. *BMC Infect Dis*. 2015;15(1):574. <https://doi.org/10.1186/s12879-015-1315-z>
69. Zheng DP, Ando T, Fankhauser RL, Beard RS, Glass RI, Monroe SS. Norovirus classification and proposed strain nomenclature. *Virology* 2006;346(2):312-23. <https://doi.org/10.1016/j.virol.2005.11.015>
70. Vinjé J. Advances in laboratory methods for detection and typing of norovirus. *J Clin Microbiol*. 2015;53(2):373-81. <https://doi.org/10.1128/JCM.01535-14>
71. Kroneman A, Vega E, Vennema H, Vinjé J, White PA, Hansman G et al. Proposal for a unified norovirus nomenclature and genotyping. *Arch Virol*. 2013;158(10):2059-68. <https://doi.org/10.1007/s00705-013-1708-5>
72. Siebenga JJ, Vennema H, Zheng DP, Vinjé J, Lee BE, Pang XL et al. Norovirus illness is a global problem: emergence and spread of norovirus GII.4 variants, 2001-2007. *J Infect Dis*. 2009;200(5):802-12. <https://doi.org/10.1086/605127>
73. Han J, Ji L, Shen Y, Wu X, Xu D, Chen L. Emergence and predominance of norovirus GII.17 in Huzhou, China, 2014-2015. *Virol J*. 2015;12(1):139. <https://doi.org/10.1186/s12985-015-0370-9>
74. Bull RA, Hansman GS, Clancy LE, Tanaka MM, Rawlinson WD, White PA. Norovirus recombination in ORF1/ORF2 overlap. *Emerg Infect Dis*. 2005;11(7):1079-85. <https://doi.org/10.3201/eid1107.041273>
75. Chhabra P, Walimbe AM, Chitambar SD. Molecular characterization of three novel intergenotype norovirus GII recombinant strains from western India. *Virus Res*. 2010;147(2):242-6. <https://doi.org/10.1016/j.virusres.2009.11.007>
76. Mahar JE, Kirkwood CD. Characterization of norovirus strains in Australian children from 2006 to 2008: prevalence of recombinant strains. *J Med Virol*. 2011;83(12):2213-9. <https://doi.org/10.1002/jmv.22215>
77. Fumian TM, Aragão GC, Mascarenhas JD, Kaiano JH, Siqueira JA, Soares LS, et al. Detection of a novel recombinant strain of norovirus in an African-descendant community from the Amazon region of Brazil in 2008. *Arch Virol*. 2012;157(12):2389-92. <https://doi.org/10.1007/s00705-012-1428-2>
78. Arana A, Cilla G, Montes M, Gomariz M, Pérez-Trallero E. Genotypes, recombinant forms, and variants of norovirus GII.4 in Gipuzkoa (Basque Country, Spain), 2009-2012. *PLoS One*. 2014;9(6):e98875. <https://doi.org/10.1371/journal.pone.0098875>
79. White PA. Evolution of norovirus. *Clin Microbiol Infect*. 2014;20(8):741-5. <https://doi.org/10.1111/1469-0691.12746>
80. Hernandez J, Silva LD, Sousa Junior EC, Lucena MS, Soares LS, Mascarenhas JD et al. Analysis of uncommon norovirus recombinants from Manaus, Amazon region, Brazil: GII.P22/GII.5, GII.P7/GII.6 and GII.Pg/GII.1. *Infect Genet Evol*. 2016;39:365-71. <https://doi.org/10.1016/j.meegid.2016.02.007>
81. Vinjé J, Green J, Lewis DC, Gallimore CI, Brown DW, Koopmans MP. Genetic polymorphism across regions of the three open reading frames of "Norwalk-like viruses". *Arch Virol*. 2000;145(2):223-41. <https://doi.org/10.1007/s007050050020>
82. Andrade JS, Rocha MS, Carvalho-Costa FA, Fioretti JM, Xavier MP, Nunes ZM et al. Noroviruses associated with outbreaks of acute gastroenteritis in the State of Rio Grande do Sul, Brazil, 2004-2011. *J Clin Virol*. 2014;61(3):345-52. <https://doi.org/10.1016/j.jcv.2014.08.024>
83. Fioretti JM, Ferreira MS, Victoria M, Vieira CB, Xavier MP, Leite JP et al. Genetic diversity of noroviruses in Brazil. *Mem Inst Oswaldo Cruz*. 2011;106(8):942-47. <https://doi.org/10.1590/S0074-02762011000800008>
84. Fioretti JM, Bello G, Rocha MS, Victoria M, Leite JP, Miagostovich MP. Temporal dynamics of norovirus GII.4 variants in Brazil between 2004 and 2012. *PLoS One*. 2014;9(3):e92988. <https://doi.org/10.1371/journal.pone.0092988>
85. Fumian TM, Leite JP, Rocha MS, Andrade JS, Fioretti JM, Assis RM et al. Performance of a one-step quantitative duplex RT-PCR for detection of rotavirus A and noroviruses GII during two periods of high viral circulation. *J Virol Methods*. 2016;(228):123-9. <https://doi.org/10.1016/j.jviromet.2015.11.008>
86. Siqueira JA, Bandeira RS, Justino MC, Linhares AC, Gabbay YB. Characterization of novel intragenotype recombination events among norovirus pandemic GII.4 variants. *Infect Genet Evol*. 2016;(44):361-6. <https://doi.org/10.1016/j.meegid.2016.07.037>
87. Lopman B, Gastañaduy P, Park GW, Hall AJ, Parashar UD, Vinjé J. Environmental transmission of norovirus gastroenteritis. *Curr Opin Virol*. 2012;2(1):96-102. <https://doi.org/10.1016/j.coviro.2011.11.005>



88. Repp KK, Keene WE. A point-source norovirus outbreak caused by exposure to fomites. *J Infect Dis*. 2012;205(11):1639-41. <https://doi.org/10.1093/infdis/jis250>
89. Petrigiani M, Beek J, Borsboom G, Richardus JH, Koopmans M. Norovirus introduction routes into nursing homes and risk factors for spread: a systematic review and meta-analysis of observational studies. *J Hosp Infect*. 2015;89(3):163-78. <https://doi.org/10.1016/j.jhin.2014.11.015>
90. Hall AJ, Wikswo ME, Pringle K, Gould LH, Parashar UD. Vital signs: foodborne norovirus outbreaks - United States, 2009-2012. *MMWR*. 2014;63(22):491-5.
91. Vivancos R, Keenan A, Sopwith W, Smith K, Quigley C, Mutton K et al. Norovirus outbreak in a cruise ship sailing around the British Isles: investigation and multi-agency management of an international outbreak. *J Infect*. 2010;60(6):478-85. <https://doi.org/10.1016/j.jinf.2010.03.018>
92. Ramani S, Atmar RL, Estes MK. Epidemiology of human noroviruses and updates on vaccine development. *Curr Opin Gastroenterol*. 2014;30(1):25-33. <https://doi.org/10.1097/MOG.000000000000022>
93. Hirneisen KA, Kniel KE. Comparing human norovirus surrogates: murine norovirus and Tulane virus. *J Food Prot*. 2013;76(7):139-43. <https://doi.org/10.4315/0362-028X.JFP-12-216>
94. Wang Q, Hirneisen KA, Markland SM, Kniel KE. Survival of murine norovirus, Tulane virus, and hepatitis A virus on alfalfa seeds and sprouts during storage and germination. *Appl Environ Microbiol*. 2013;79(22):7021-27. <https://doi.org/10.1128/AEM.01704-13>
95. Baert L, Uyttendaele M, Stals A, VAN Coillie E, Dierick K, Debevere J et al. Reported foodborne outbreaks due to noroviruses in Belgium: the link between food and patient investigations in an international context. *Epidemiol Infect*. 2009;137(3):316-25. <https://doi.org/10.1017/S0950268808001830>
96. Dolin R, Blacklow NR, DuPont H, Buscho RF, Wyatt RG, Kasel JA et al. Biological properties of Norwalk agent of acute infectious nonbacterial gastroenteritis. *Proc Soc Exp Biol Med*. 1972;140:578-83. <https://doi.org/10.3181/00379727-140-36508>
97. Green KY. *Caliciviridae: the noroviruses*. Philadelphia: Lippincott-Raven Publishers PA;2007.
98. Mormann S, Dabisch M, Becker B. Effects of technological processes on the tenacity and inactivation of norovirus genogroup II in experimentally contaminated foods. *Appl Environ Microbiol*. 2010;76(2):536-45. <https://doi.org/10.1128/AEM.01797-09>
99. Kingsley DH. High pressure processing and its application to the challenge of virus-contaminated foods. *Food Environ Virol*. 2013;5(1):1-12. <https://doi.org/10.1007/s12560-012-9094-9>
100. Baert L, Debevere J, Uyttendaele M. The efficacy of preservation methods to inactivate foodborne viruses. *Int J Food Microbiol*. 2009;131(2-3):83-94. <https://doi.org/10.1016/j.ijfoodmicro.2009.03.007>
101. Wobus CE, Thackray LB, Virgin HW 4th. Murine norovirus: a model system to study norovirus biology and pathogenesis. *J Virol*. 2006;80(11):5104-12. <https://doi.org/10.1128/JVI.02346-05>
102. Takahashi M, Takahashi H, Kuda T, Kimura B. Viability and heat resistance of murine norovirus on bread. *Int J Food Microbiol*. 2016;216:127-31. <https://doi.org/10.1016/j.ijfoodmicro.2015.09.018>
103. Rutjes SA, Lodder-Verschoor F, Poel WH, Duijnhoven YT, Roda Husman AM. Detection of noroviruses in foods: a study on virus extraction procedures in foods implicated in outbreaks of human gastroenteritis. *J Food Prot*. 2006;69(8):1949-56. <https://doi.org/10.4315/0362-028X-69.8.1949>
104. Farkas T, Sestak K, Wei C, Jiang X. Characterization of a rhesus monkey calicivirus representing a new genus of Caliciviridae. *J Virol*. 2008;82(11):5408-16. <https://doi.org/10.1128/JVI.00070-08>
105. Matthews JE, Dickey BW, Miller RD, Felzer JR, Dawson BP, Lee AS et al. The epidemiology of published norovirus outbreaks: a review of risk factors associated with attack rate and genogroup. *Epidemiol Infect*. 2012;140(7):1161-72. <https://doi.org/10.1017/S0950268812000234>
106. Xerry J, Gallimore CI, Iturriza-Gómara M, Gray JJ. Tracking the transmission routes of genogroup II noroviruses in suspected food-borne or environmental outbreaks of gastroenteritis through sequence analysis of the P2 domain. *J Med Virol*. 2009;81(7):1298-304. <https://doi.org/10.1002/jmv.21517>
107. Anderson AD, Garrett VD, Sobel J, Monroe SS, Fankhauser RL, Schwab KJ et al. Multistate outbreak of Norwalk-like virus gastroenteritis associated with a common caterer. *Am J Epidemiol*. 2001;154(11):1013-19. <https://doi.org/10.1093/aje/154.11.1013>
108. Grove SF, Suriyanarayanan A, Puli B, Zhao H, Li M, Li D et al. Norovirus cross-contamination during preparation of fresh produce. *Int J Food Microbiol*. 2015;198:43-9. <https://doi.org/10.1016/j.ijfoodmicro.2014.12.023>
109. Sair AI, D'Souza DH, Moe CL, Jaykus LA. Improved detection of human enteric viruses in foods by RT-PCR. *J Virol Methods*. 2002;100(1-2):57-69. [https://doi.org/10.1016/S0166-0934\(01\)00397-4](https://doi.org/10.1016/S0166-0934(01)00397-4)
110. Posada-Izquierdo GD, Pérez-Rodríguez F, López-Gálvez F, Allende A, Gil MI, Zurera G. Modeling growth of *Escherichia coli* O157:H7 in fresh-cut lettuce treated with neutral electrolyzed water and under modified atmosphere packaging. *Int J Food Microbiol*. 2014;177(2):1-8. <https://doi.org/10.1016/j.ijfoodmicro.2013.12.025>
111. Daniels NA, Bergmire-Sweat DA, Schwab KJ, Hendricks KA, Reddy S, Rowe SM et al. A foodborne outbreak of gastroenteritis associated with Norwalk-like viruses: first molecular traceback to deli sandwiches contaminated during preparation. *J Infect Dis*. 2000;181(4):1467-70. <https://doi.org/10.1086/315365>



112. Stals A, Jacxsens L, Baert L, Van Coillie E, Uyttendaele M. A quantitative exposure model simulating human norovirus transmission during preparation of deli sandwiches. *Int J Food Microbiol.* 2015;196:126-36. <https://doi.org/10.1016/j.ijfoodmicro.2014.12.004>
113. Kobayashi S, Natori K, Takeda N, Sakae K. Immunomagnetic capture rt-PCR for detection of norovirus from foods implicated in a foodborne outbreak. *Microbiol Immunol.* 2004;48(3):201-4. <https://doi.org/10.1111/j.1348-0421.2004.tb03506.x>
114. Baert L, Uyttendaele M, Debevere J. Evaluation of viral extraction methods on a broad range of Ready-To-Eat foods with conventional and real-time RT-PCR for Norovirus GII detection. *Int J Food Microbiol.* 2008;123(1-2):101-8. <https://doi.org/10.1016/j.ijfoodmicro.2007.12.020>
115. Doyle MP, Erickson MC. Summer meeting 2007: the problems with fresh produce: an overview. *J Appl Microbiol.* 2008;105:317-30. <https://doi.org/10.1111/j.1365-2672.2008.03746.x>
116. Heaton JC, Jones K. Microbial contamination of fruit and vegetables and the behaviour of enteropathogens in the phyllosphere: a review. *J Appl Microbiol.* 2008;104(3):613-26. <https://doi.org/10.1111/j.1365-2672.2007.03587.x>
117. Sarvikivi E, Roivainen M, Maunula L, Niskanen T, Korhonen T, Lappalainen M et al. Multiple norovirus outbreaks linked to imported frozen raspberries. *Epidemiol Infect.* 2012;140(2):260-7. <https://doi.org/10.1017/S0950268811000379>
118. El-Senousy WM, Costafreda MI, Pintó RM, Bosch A. Method validation for norovirus detection in naturally contaminated irrigation water and fresh produce. *Int J Food Microbiol.* 2013;167(1):74-9. <https://doi.org/10.1016/j.ijfoodmicro.2013.06.023>
119. Dicaprio E, Ma Y, Purgianto A, Hughes J, Li J. Internalization and dissemination of human norovirus and animal caliciviruses in hydroponically grown romaine lettuce. *Appl Environ Microbiol.* 2012;78(17):6143-52. <https://doi.org/10.1128/AEM.01081-12>
120. Erickson MC. Internalization of fresh produce by foodborne pathogens. *Annu Rev Food Sci Technol.* 2012;3(1):283-310. <https://doi.org/10.1146/annurev-food-022811-101211>
121. Jaykus LA, Escudero-Abarca B. Human pathogenic viruses in food. In: Juneja VK, Sofos JN, editors. *Pathogens and toxins in foods: challenges and interventions.* Washington DC: ASM Press; 2010. p. 218-32.
122. Li J, Predmore A, Divers E, Lou F. New interventions against human norovirus: progress, opportunities, and challenges. *Annu Rev Food Sci Technol.* 2012;3:331-52. <https://doi.org/10.1146/annurev-food-022811-101234>
123. European Food Safety Authority, European Centre for Disease Prevention and Control. *The European Union summary report on trends and sources of zoonoses, zoonotic agents and foodborne outbreaks in 2013.* EFSA J. 2015;13(1):3991. <https://doi.org/10.2903/j.efsa.2015.3991>
124. Carducci A, Caponi E, Ciurli A, Verani M. Possible internalization of an enterovirus in hydroponically grown lettuce. *Int J Environ Res Public Health.* 2015;12(7):8214-27. <https://doi.org/10.3390/ijerph120708214>
125. Le Guyader FS, Bon F, DeMedici D, Parnaudeau S, Bertone A, Crudeli S et al. Detection of multiple noroviruses associated with an international gastroenteritis outbreak linked to oyster consumption. *J Clin Microbiol.* 2006;44(11):3878-82. <https://doi.org/10.1128/JCM.01327-06>
126. Burkhardt W 3rd, Calci KR. Selective accumulation may account for shellfish-associated viral illness. *Appl Environ Microbiol.* 2000;66(4):1375-8. <https://doi.org/10.1128/AEM.66.4.1375-1378.2000>
127. Le Guyader FS, Atmar RL, Le Pendu J. Transmission of viruses through shellfish: when specific ligands come into play. *Curr Opin Virol.* 2012;2(1):103-10. <https://doi.org/10.1016/j.coviro.2011.10.029>
128. Alfano-Sobsey E, Sweat D, Hall A, Breedlove F, Rodriguez R, Greene S et al. Norovirus outbreak associated with undercooked oysters and secondary household transmission. *Epidemiol Infect.* 2012;140(2):276-82. <https://doi.org/10.1017/S0950268811000665>
129. Webby RJ, Carville KS, Kirk MD, Greening G, Ratcliff RM, Crerar SK et al. Internationally distributed frozen oyster meat causing multiple outbreaks of norovirus infection in Australia. *Clin Infect Dis.* 2007;44(8):1026-31. <https://doi.org/10.1086/512807>
130. Centers for Disease Control and Prevention (CDC). Notes from the field: norovirus infections associated with frozen raw oysters - Washington, 2011. *MMWR.* 2012;61(6):110.
131. Terio V, Martella V, Moschidou P, Di Pinto P, Tantillo G, Buonavoglia C. Norovirus in retail shellfish. *Food Microbiol.* 2010;27(1):29-32. <https://doi.org/10.1016/j.fm.2009.07.005>
132. DePaola A, Jones JL, Woods J, Burkhardt W 3rd, Calci KR, Krantz JA et al. Bacterial and viral pathogens in live oysters: 2007 United States market survey. *Appl Environ Microbiol.* 2010;76(9):2754-68. <https://doi.org/10.1128/AEM.02590-09>
133. Souza DS, Piazza RS, Pilotto MR, Nascimento MA, Moresco V, Taniguchi S et al. Virus, protozoa and organic compounds decay in depurated oysters. *Int J Food Microbiol.* 2013;167(3):337-45. <https://doi.org/10.1016/j.ijfoodmicro.2013.09.019>
134. Lowther JA, Gustar NE, Powell AL, Hartnell RE, Lees DN. Two-year systematic study to assess norovirus contamination in oysters from commercial harvesting areas in the United Kingdom. *Appl Environ Microbiol.* 2012;78(16):5812-7. <https://doi.org/10.1128/AEM.01046-12>
135. Baker K, Morris J, McCarthy N, Saldana L, Lowther J, Collinson A et al. An outbreak of norovirus infection linked to oyster consumption at a UK restaurant, February 2010. *J Public Health (Oxf).* 2011;33(2):205-11. <https://doi.org/10.1093/pubmed/fdq089>



136. Souza DS, Ramos AP, Nunes FF, Moresco V, Taniguchi S, Leal DA et al. Evaluation of tropical water sources and mollusks in southern Brazil using microbiological, biochemical, and chemical parameters. *Ecotoxicol Environ Saf.* 2012;76(2):153-61. <https://doi.org/10.1016/j.ecoenv.2011.09.018>
137. Stals A, Baert L, De Keuckelaere A, Van Coillie E, Uyttendaele M. Evaluation of a norovirus detection methodology for ready-to-eat foods. *Int J Food Microbiol.* 2011;145(2-3):420-5. <https://doi.org/10.1016/j.ijfoodmicro.2011.01.013>
138. Malek M, Barzilay E, Kramer A, Camp B, Jaykus LA, Escudero-Abarca B et al. Outbreak of norovirus infection among river rafters associated with packaged delicatessen meat, Grand Canyon, 2005. *Clin Infect Dis.* 2009;48(1):31-7. <https://doi.org/10.1086/594118>
139. Aragão GC, Mascarenhas JD, Kaiano JH, de Lucena MS, Siqueira JA, Fumian TM et al. Norovirus diversity in diarrheic children from an African-descendant settlement in Belém, Northern Brazil. *PLoS One.* 2013;8(2):e56608. <https://doi.org/10.1371/journal.pone.0056608>
140. Moore MD, Goulter RM, Jaykus LA. Human norovirus as a foodborne pathogen: challenges and developments. *Annu Rev Food Sci Technol.* 2015;6(1):411-33. <https://doi.org/10.1146/annurev-food-022814-015643>
141. Baert L, Mattison K, Loisy-Hamon F, Harlow J, Martyres A, Lebeau B et al. Review: norovirus prevalence in Belgian, Canadian and French fresh produce: a threat to human health? *Int J Food Microbiol.* 2011;151(3):261-9. <https://doi.org/10.1016/j.ijfoodmicro.2011.09.013>
142. Boxman IL, Verhoef L, Dijkman R, Hägele G, Te Loeke NA, Koopmans M. Year-round prevalence of norovirus in the environment of catering companies without a recently reported outbreak of gastroenteritis. *Appl Environ Microbiol.* 2011;77(9):2968-74. <https://doi.org/10.1128/AEM.02354-10>
143. Schwab KJ, McDevitt JJ. Development of a PCR-enzyme immunoassay oligoprobe detection method for *Toxoplasma gondii* oocysts, incorporating PCR controls. *Appl Environ Microbiol.* 2003;69(10):5819-25. <https://doi.org/10.1128/AEM.69.10.5819-5825.2003>
144. Demeke T, Jenkins GR. Influence of DNA extraction methods, PCR inhibitors and quantification methods on real-time PCR assay of biotechnology-derived traits. *Anal Bioanal Chem.* 2010;396(6):1977-90. <https://doi.org/10.1007/s00216-009-3150-9>
145. Summa M, von Bonsdorff CH, Maunula L. Evaluation of four virus recovery methods for detecting noroviruses on fresh lettuce, sliced ham, and frozen raspberries. *J Virol Methods.* 2012;183(2):154-60. <https://doi.org/10.1016/j.jviromet.2012.04.006>
146. Katayama H, Shimasaki A, Ohgaki S. Development of a virus concentration method and its application to detection of enterovirus and norwalk virus from coastal seawater. *Appl Environ Microbiol.* 2002;68(3):1033-9. <https://doi.org/10.1128/AEM.68.3.1033-1039.2002>
147. Fumian TM, Leite JP, Marin VA, Miagostovich MP. A rapid procedure for detecting noroviruses from cheese and fresh lettuce. *J Virol Methods.* 2009;155(1):39-43. <https://doi.org/10.1016/j.jviromet.2008.09.026>
148. Brandão ML, Almeida DO, Bispo FC, Bricio SM, Marin VA, Miagostovich MP. Assessment of microbiological contamination of fresh, minimally processed, and ready-to-eat lettuces (*Lactuca sativa*), Rio de Janeiro State, Brazil. *J Food Sci.* 2014;79(5):M961-6. <https://doi.org/10.1111/1750-3841.12459>
149. Corrêa AA, Miagostovich MP. Optimization of an adsorption-elution method with a negatively charged membrane to recover norovirus from lettuce. *Food Environ. Virol* 2013;5(3):144-9. <https://doi.org/10.1007/s12560-013-9113-5>
150. Calgua B, Mengewein A, Grunert A, Bofill-Mas S, Clemente-Casares P, Hundesa A et al. Development and application of a one-step low cost procedure to concentrate viruses from seawater samples. *J Virol Methods.* 2008;153(2):79-83. <https://doi.org/10.1016/j.jviromet.2008.08.003>
151. Melgaço FG, Victoria M, Corrêa AA, Ganime AC, Malta FC, Brandão ML et al. Virus recovering from strawberries: evaluation of a skimmed milk organic flocculation method for assessment of microbiological contamination. *Int J Food Microbiol.* 2016;217:14-9. <https://doi.org/10.1016/j.ijfoodmicro.2015.10.005>
152. Schwab KJ, Neill FH, Fankhauser RL, Daniels NA, Monroe SS, Bergmire-Sweat DA et al. Development of methods to detect "Norwalk-like viruses" (NLVs) and hepatitis A virus in delicatessen foods: application to a food-borne NLV outbreak. *Appl Environ Microbiol.* 2000;66(1):213-8. <https://doi.org/10.1128/AEM.66.1.213-218.2000>
153. Boxman IL, Tilburg JJ, te Loeke NA, Vennema H, Boer E, Koopmans M. An efficient and rapid method for recovery of norovirus from food associated with outbreaks of gastroenteritis. *J Food Prot.* 2007;70(2):504-8. <https://doi.org/10.4315/0362-028X-70.2.504>
154. Höhne M, Schreier E. Detection and characterization of norovirus outbreaks in Germany: application of a one-tube RT-PCR using a fluorogenic real-time detection system. *J Med Virol.* 2004;72(2):312-9. <https://doi.org/10.1002/jmv.10573>
155. Bustin SA, Nolan T. Pitfalls of quantitative real-time reverse-transcription polymerase chain reaction. *J Biomol Tech.* 2004;15(3):155-66.
156. Barker J, Vipond IB, Bloomfield SF. Effects of cleaning and disinfection in reducing the spread of Norovirus contamination via environmental surfaces. *J Hosp Infect.* 2004;58(1):42-9. <https://doi.org/10.1016/j.jhin.2004.04.021>
157. Knight A, Li D, Uyttendaele M, Jaykus LA. A critical review of methods for detecting human noroviruses and predicting their infectivity. *Crit Rev Microbiol.* 2013;39(3):295-309. <https://doi.org/10.3109/1040841X.2012.709820>



158. Escudero-Abarca BI, Suh SH, Moore MD, Dwivedi HP, Jaykus LA. Selection, characterization and application of nucleic acid aptamers for the capture and detection of human norovirus strains. *PLoS One*. 2014;9(9):e106805. <https://doi.org/10.1371/journal.pone.0106805>
159. Prevost B, Lucas FS, Ambert-Balay K, Pothier P, Moulin L, Wurtzer S. Deciphering the diversities of astroviruses and noroviruses in wastewater treatment plant effluents by a high-throughput sequencing method. *Appl Environ Microbiol*. 2015;81(20):7215-22. <https://doi.org/10.1128/AEM.02076-15>
160. Sánchez G, Elizaquível P, Aznar R. A single method for recovery and concentration of enteric viruses and bacteria from fresh-cut vegetables. *Int J Food Microbiol*. 2012;152(1-2):9-13. <https://doi.org/10.1016/j.ijfoodmicro.2011.10.002>
161. Bae J, Schwab KJ. Evaluation of murine norovirus, feline calicivirus, poliovirus, and MS2 as surrogates for human norovirus in a model of viral persistence in surface water and groundwater. *Appl Environ Microbiol*. 2008;74(2):477-84. <https://doi.org/10.1128/AEM.02095-06>
162. Hennechart-Collette C, Martin-Latit S, Guillier L, Perelle S. Multiplex real-time RT-qPCR for the detection of Norovirus in bottled and tap water using murine norovirus as a process control. *J Appl Microbiol*. 2014;116(1):179-90. <https://doi.org/10.1111/jam.12345>
163. Martin-Latit S, Hennechart-Collette C, Guillier L, Perelle S. Duplex RT-qPCR for the detection of hepatitis E virus in water, using a process control. *Int J Food Microbiol*. 2012;157(2):167-73. <https://doi.org/10.1016/j.ijfoodmicro.2012.05.001>
164. Stals A, Baert L, Van Coillie E, Uyttendaele M. Evaluation of a norovirus detection methodology for soft red fruits. *Food Microbiol*. 2011;28(1):52-8. <https://doi.org/10.1016/j.fm.2010.08.004>
165. Costafreda MI, Bosch A, Pintó RM. Development, evaluation, and standardization of a real-time TaqMan reverse transcription-PCR assay for quantification of hepatitis A virus in clinical and shellfish samples. *Appl Environ Microbiol*. 2006;72(6):3846-55. <https://doi.org/10.1128/AEM.02660-05>
166. Uhrbrand K, Myrnel M, Maunula L, Vainio K, Trebbien R, Nørrung B et al. Evaluation of a rapid method for recovery of norovirus and hepatitis A virus from oysters and blue mussels. *J Virol Methods*. 2010;169(1):70-8. <https://doi.org/10.1016/j.jviromet.2010.06.019>
167. Rajal VB, McSwain BS, Thompson DE, Leutenegger CM, Kildare BJ, Wuertz S. Validation of hollow fiber ultrafiltration and real-time PCR using bacteriophage PP7 as surrogate for the quantification of viruses from water samples. *Water Res*. 2007;41(7):1411-22. <https://doi.org/10.1016/j.watres.2006.12.034>
168. Noel JS, Ando T, Leite JP, Green KY, Dingle KE, Estes MK et al. Correlation of patient immune responses with genetically characterized small round-structured viruses involved in outbreaks of nonbacterial acute gastroenteritis in the United States, 1990 to 1995. *J Med Virol*. 1997;53(4):372-83. [https://doi.org/10.1002/\(SICI\)1096-9071\(199712\)53:4<372::AID-JMV10>3.0.CO;2-H](https://doi.org/10.1002/(SICI)1096-9071(199712)53:4<372::AID-JMV10>3.0.CO;2-H)
169. Kojima S, Kageyama T, Fukushi S, Hoshino FB, Shinohara M, Uchida K et al. Genogroup-specific PCR primers for detection of Norwalk-like viruses. *J Virol Methods*. 2002;100(1-2):107-14. [https://doi.org/10.1016/S0166-0934\(01\)00404-9](https://doi.org/10.1016/S0166-0934(01)00404-9)
170. Vinjé J, Hamidjaja RA, Sobsey MD. Development and application of a capsid VP1 (region D) based reverse transcription PCR assay for genotyping of genogroup I and II noroviruses. *J Virol Methods*. 2004;116(2):109-17. <https://doi.org/10.1016/j.jviromet.2003.11.001>
171. Vega E, Barclay L, Gregoricus N, Williams K, Lee D, Vinjé J. Novel surveillance network for norovirus gastroenteritis outbreaks, United States. *Emerg Infect Dis*. 2011;17(8):1389-95.
172. Kroneman A, Vennema H, Deforche K, Avoort H, Peñaranda S, Oberste MS et al. An automated genotyping tool for enteroviruses and noroviruses. *J Clin Virol*. 2011;51(2):121-5. <https://doi.org/10.1016/j.jcv.2011.03.006>
173. Franz E, Tromp SO, Rijgersberg H, Fels-Klerx HJ. Quantitative microbial risk assessment for *Escherichia coli* O157:H7, salmonella, and *Listeria monocytogenes* in leafy green vegetables consumed at salad bars. *J Food Prot*. 2010;73(2):274-85. <https://doi.org/10.4315/0362-028X-73.2.274>
174. Carrasco E, Pérez-Rodríguez F, Valero A, Garcia-Gimeno R, Zurera G. Risk assessment and management of *Listeria monocytogenes* in Ready-to-Eat lettuce salads. *Compr Rev Food Sci Food Saf*. 2010;9(5):498-512. <https://doi.org/10.1111/j.1541-4337.2010.00123.x>
175. Sant'Ana AS, Franco BD, Schaffner DW. Risk of infection with *Salmonella* and *Listeria monocytogenes* due to consumption of ready-to-eat leafy vegetables in Brazil. *Food Control*. 2014;42:1-8. <https://doi.org/10.1016/j.foodcont.2014.01.028>
176. Aström J, Petterson S, Bergstedt O, Pettersson TJ, Stenström TA. Evaluation of the microbial risk reduction due to selective closure of the raw water intake before drinking water treatment. *J Water Health*. 2007;5 Suppl 1:81-97. <https://doi.org/10.2166/wh.2007.139>
177. Schoen ME, Ashbolt NJ. Assessing pathogen risk to swimmers at non-sewage impacted recreational beaches. *Environ Sci Technol*. 2010;44(7):2286-91. <https://doi.org/10.1021/es903523q>
178. Soller JA, Schoen ME, Bartrand T, Ravenscroft JE, Ashbolt NJ. Estimated human health risks from exposure to recreational waters impacted by human and non-human sources of faecal contamination. *Water Res* 2010;44(16):4674-91. <https://doi.org/10.1016/j.watres.2010.06.049>
179. Hamilton AJ, Stagnitti F, Premier R, Boland AM, Hale G. Quantitative microbial risk assessment models for consumption of raw vegetables irrigated with reclaimed water. *Appl Environ Microbiol*. 2006;72(5):3284-90. <https://doi.org/10.1128/AEM.72.5.3284-3290.2006>



180. Mara D, Sleight A. Estimation of norovirus infection risks to consumers of wastewater-irrigated food crops eaten raw. *J Water Health*. 2010;8(1):39-43. <https://doi.org/10.2166/wh.2009.140>
181. Kotwal G, Cannon J. Norovirus cross-contamination associated with bare hands and gloves during produce handling. In: IAFP 2013 Annual Meeting; 29-31 jul 2013[acesso 8 dez 2014]; Des Moines, USA. Disponível em: <https://iafp.confex.com/iafp/2013/webprogram/Paper4776.html>
182. De Keuckelaere A, Li D, Deliens B, Stals A, Uyttendaele M. Batch testing for noroviruses in frozen raspberries. *Int J Food Microbiol*. 2015;192:43-50. <https://doi.org/10.1016/j.ijfoodmicro.2014.09.024>
183. Barclay L, Park GW, Vega E, Hall A, Parashar U, Vinjé J et al. Infection control for norovirus. *Clin Microbiol Infect*. 2014;20(8):731-40. <https://doi.org/10.1111/1469-0691.12674>
184. Patel MM, Hall AJ, Vinjé J, Parashar UD. Noroviruses: a comprehensive review. *J Clin Virol*. 2009;44(1):1-8. <https://doi.org/10.1016/j.jcv.2008.10.009>
185. Centers for Disease Control and Prevention (CDC). Surveillance for foodborne disease outbreaks - United States, 2008. *MMWR*. 2011;60(35):1197-202.
186. Zhang X, Buehner NA, Hutson AM, Estes MK, Mason HS. Tomato is a highly effective vehicle for expression and oral immunization with Norwalk virus capsid protein. *Plant Biotechnol J*. 2006;4(4):419-32. <https://doi.org/10.1111/j.1467-7652.2006.00191.x>
187. El-Kamary SS, Pasetti MF, Mendelman PM, Frey SE, Bernstein DI, Treanor JJ et al. Adjuvanted intranasal Norwalk virus-like particle vaccine elicits antibodies and antibody-secreting cells that express homing receptors for mucosal and peripheral lymphoid tissues. *J Infect Dis*. 2010;202(11):1649-58. <https://doi.org/10.1086/657087>
188. Atmar RL, Bernstein DI, Harro CD, Al-Ibrahim MS, Chen WH, Ferreira J, et al. Norovirus vaccine against experimental human Norwalk Virus illness. *N Engl J Med*. 2011;365(23):2178-87. <https://doi.org/10.1056/NEJMoa1101245>
189. Treanor JJ, Atmar RL, Frey SE, Gormley R, Chen WH, Ferreira J, et al. A novel intramuscular bivalent norovirus virus-like particle vaccine candidate - reactogenicity, safety, and immunogenicity in a phase 1 trial in healthy adults. *J Infect Dis*. 2014;210(11):1763-71. <https://doi.org/10.1093/infdis/jiu337>
190. Bernstein DI, Atmar RL, Lyon GM, Treanor JJ, Chen WH, Jiang X et al. Norovirus vaccine against experimental human GII.4 virus illness: a challenge study in healthy adults. *J Infect Dis*. 2015;211(6):870-8. <https://doi.org/10.1093/infdis/jiu497>
191. Vinjé J. A norovirus vaccine on the horizon? *J Infect Dis*. 2010;202(11):1623-5. <https://doi.org/10.1086/657088>

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Conflict of Interest

Authors have no potential conflict of interest to declare, related to this study's political or financial peers and institutions.



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